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OF

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FOR

MAMMALIAN TRANSFORMING GROWTH FACTOR BETA - 9

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This claims the benefit of U.S. Provisional Application Serial No. 60/100,706 filed September 17, 1998.

10 BACKGROUND OF THE INVENTION

Proper control of the opposing processes of cell proliferation versus terminal differentiation and apoptotic programmed cell death is an important aspect of normal development and homeostasis, Raff, M.C., *Cell*, 15 86:173-175 (1996), and has been found to be altered in many human diseases. See, for example, Sawyers, C.L. et al., *Cell*, 64:337-350 (1991); Meyaard, L. et al., *Science*, 257:217-219 (1992); Guo, Q. et al., *Nature Med.*, 4:957-962 (1998); Barinaga, M. *Science*, 273:735-737 (1996); Solary, 20 E. et al., *Eur. Respir. J.*, 9:1293-1305 (1996); Hamet, P. et al., *J. Hypertension*, 14:S65-S70, (1996); Roy, N. et al. *Cell*, 80:167-178 (1995); and Ambrosini, G., *Nature Med.*, 8:917-921 (1997). Much progress has been made towards understanding the regulation of this balance. For 25 example, signaling cascades have been elucidated through which extracellular stimuli, such as growth factors, peptide hormones, and cell-cell interactions control the commitment of precursor cells to specific lineages and their subsequent proliferative expansion, Morrison, S.J. 30 et al., *Cell*, 88:287-298 (1997). Further, it has been found that cell cycle exit and terminal differentiation are coupled in most cell types. See, for example, Coppola, J.A. et al. *Nature*, 320:760-763 (1986); Freytag, S.O., *Mol. Cell. Biol.* 8:1614-1624 (1988); Lee, E.Y. et al., 35 *Genes Dev.*, 8:2008-2021 (1994); Morgenbesser, S.D., et al., *Nature*, 371:72-74 (1994); Casaccia-Bonnet, P. et al., *Genes Dev.*, 11:2335-2346 (1996); Zacksenhaus, E. et

al., *Genes Dev.*, 10:3051-3064 (1996); and Zhang, P. et al., *Nature*, 387:151-158 (1997). Apoptosis (programmed cell death) also plays an important role in many developmental and homeostatic processes, Raff, M.C.,
 5 *Nature*, 356:397-400 (1992), and is often coordinately regulated with terminal differentiation, Jacobsen, K.A. et al., *Blood*, 84:2784-2794 (1994); Yan, Y. et al., *Genes Dev.*, 11:973-983 (1997). Hence, it appears that the cell
 10 type of individual lineages, tissues, organs, or even entire multicellular organisms is the result of a finely tuned balance between increased cell production due to proliferation, and decreased numbers of cells resulting from terminal differentiation and apoptosis. This balance is most likely regulated coordinately by the convergence
 15 of multiple regulatory pathways. The identification of novel members of such networks can provide important insights into both normal cellular processes as well as the etiology and treatment of human disease states.

20 Interleukin 17 (IL-17) is a cytokine which has been implicated as an important regulator of the immune system, Spriggs, M.K., *J. Clinical Immunology*, 17:366-369 (1997), Broxmeyer, H.E., *J. Experimental Medicine*, 183:2411-2415 (1996), Yao, Z., et al., *J. Immunology*,
 25 155:5483-5486 (1995), Yao, Z., et al., *Immunity*, 3:811-821 (1995). Human IL-17 is almost exclusively produced by activated CD4+ memory T cells (however, in mice, CD4-/CD8- T cells also express IL-17), Aarvak, T., et al., *J. Immunology*, 162:1246-1251 (1999), Kennedy, J., et al., *J. Interferon Cytokine Research*, 16:611-617 (1996). In
 30 contrast, the IL-17 receptor (IL-17R) appears to be ubiquitously expressed, Yao, Z., et al., *Immunity*, 3:811-821 (1995). IL-17 induces the secretion of IL-6, IL-8, monocyte chemotactic peptide-1 and G-CSF from a variety of
 35 different stromal cell types, but has no effect on cytokine production by lymphoid cells, Teunissen, M.B.M., *J. Investigative*

Dermatology, 111:645-649 (1998), Jovanovic, D.V., et al.,
 J. Immunology, 160:3513-3521 (1998), Chabaud, M., et al.,
 J. Immunology, 161:409-414 (1998), Cai, X.-Y., et al.,
 Immunology Letters, 62:51-58 (1998), Fossiez, F., et al.,
 5 J. Experimental Medicine, 183:2593-2603 (1996). IL-17
 also enhances the expression of ICAM-1 adhesion molecules
 on fibroblasts, and can stimulate granulopoiesis,
 Schwarzenberger P., et al., J. Immunology,
 161:6383-9 (1998). . Taken together, these observations
 10 have suggested that IL-17 functions as a pro-inflammatory
 cytokine. IL-17 also promotes dendritic cell
 differentiation, osteoclastogenesis, can induce nitric
 oxide production in human osteoarthritis cartilage, and is
 present in synovial fluids from patients with rheumatoid
 15 arthritis, Antonysamy, M.A., et al., J. Immunology,
 162:577-584 (1999), Kotake, S., et al., J. Clinical
 Investigation, 103:1345-1352, (1999), Attur, M.G., et al.,
 Arthritis & Rheumatism, 40:1050-1053 (1997). Blocking IL-
 17 with a soluble IL-17R protein was found to suppress
 20 cardiac allograft rejection, which correlated with
 increased IL-17 mRNA in kidney biopsies from humans
 undergoing renal allograft rejection, Antonysamy, M.A., et
 al., J. Immunology, 162:577-584 (1999). Increased IL-17
 mRNA expression is also observed in humans with multiple
 25 sclerosis, Matusevicius, D. et al., Multiple Sclerosis,
 5:101-104 (1999). Further, IL-17 can promote
 tumorigenicity of human cervical tumors in nude mice,
 Tartour, E. et al., Cancer Res., 59:3698-36704 (1999).
 Hence, IL-17 appears to play an essential role in
 30 regulating the immune system and inflammatory processes.

Thus, there is a continuing need to discover new
 proteins involved with proliferation, differentiation, and
 35 apoptotic pathways. The in vivo activities of both
 inducers and inhibitors of these pathways illustrates the
 enormous clinical potential of, and need for, novel

proliferation, differentiation, and apoptotic proteins, their agonists and antagonists. There is also a need to discover new agents which have anti-viral activity.

5

SUMMARY OF THE INVENTION

The present invention addresses this need by providing a novel anti-viral polypeptide called
10 transforming growth factor beta-9, hereinafter referred to as Ztgf β -9, and related compositions and methods. This polypeptide has anti-viral activity as disclosed in Example 10 below. It may also be used to regulate the proliferation, differentiation and apoptosis of neurons
15 glial cells, lymphocytes, hematopoietic cells and stromal cells.

Thus, one aspect of the present invention provides for an isolated Ztgf β -9 polypeptide and
20 polynucleotide. The human sequences are defined by SEQ ID NOs: 1 and 2.

The nucleotide sequence of SEQ ID NO:1 contains an open reading frame encoding a polypeptide of about 202
25 amino acids with the initial Met as shown in SEQ ID NO:1 and SEQ ID NO:2. A predicted signal sequence is comprised of amino acid residues 1, a methionine extending to and includes amino acid residue 15, an alanine. Thus a mature sequence excluding the signal sequence extends from amino
30 acid residue 16, an alanine, to and including amino acid residue 202—a proline, of SEQ ID NO:2. This mature sequence is also represented by SEQ ID NO:3. In an alternative embodiment the signal sequence extends to and includes amino acid residue 16, an alanine. This produces
35 a mature sequence which extends from amino acid 17, a glycine, to and including amino acid residue 202, a proline, of SEQ ID NO:2. This mature sequence is also

represented by SEQ ID NO:4. In another alternative embodiment, the signal sequence extends to and includes amino acid residue 17, a glycine. This results in a mature sequence which extends from amino acid residue 18, an alanine, to and including amino acid residue 202, a proline, of SEQ ID NO:2. This mature sequence is further represented by SEQ ID NO:5. Another variant of Ztgfb-9 is disclosed by SEQ ID NOs: 16 and 17. The mature sequence extends from amino acid residue 23, an alanine, to and including amino acid residue 209, a proline. The mature sequence is also defined by SEQ ID NO:18.

Murine Ztgfb-9 is defined by SEQ ID NOs: 8 and 9. The signal sequence extends from the methionine at position 1 through the alanine at position 22. Thus the mature sequence extends from the alanine at position 23 of SEQ ID NO:9 through the arginine at position 205. The mature sequence is further represented by SEQ ID NO:12.

An additional embodiment of the present invention relates to a peptide or polypeptide which has the amino acid sequence of an epitope-bearing portion of a Ztgfb-9 polypeptide having an amino acid sequence described above. Peptides or polypeptides having the amino acid sequence of an epitope-bearing portion of a Ztgfb-9 polypeptide of the present invention include portions of such polypeptides with at least nine, preferably at least 15 and more preferably at least 30 to 50 amino acids, although epitope-bearing polypeptides of any length up to and including the entire amino acid sequence of a polypeptide of the present invention described above are also included in the present invention. Examples of such epitope-bearing polypeptides are SEQ ID NOs: 13, 14, 15, 19, 20, 21 and 22. Also claimed are any of these polypeptides that are fused to another polypeptide or carrier molecule. Also claimed is an isolated nucleic acid

which encodes an epitope-bearing portion of a Ztgf β -9 polypeptide.

The present invention is further comprised of an
5 isolated peptide or polypeptide of the above-described
peptides or polypeptides having an amino acid sequence
modified by addition, deletion and/or replacement of one
or more amino acid residues and which maintains the
biological activity of said peptide or polypeptide.

10

Within a further aspect of the invention there
is provided a chimeric polypeptide consisting essentially
of a first portion and a second portion joined by a
peptide bond. The first portion of the chimeric
15 polypeptide consists essentially of (a) a Ztgf β -9
polypeptide as described above (b) allelic variants of the
polypeptides described above. The second portion of the
chimeric polypeptide consists essentially of another
polypeptide such as an affinity tag. Within one
20 embodiment the affinity tag is an immunoglobulin F_C
polypeptide. The invention also provides expression
vectors encoding the chimeric polypeptides and host cells
transfected to produce the chimeric polypeptides.

25 Another aspect of the present invention provides
for isolated nucleic acid molecules comprising a
polynucleotide selected from the group consisting of: (a)
a nucleotide sequence encoding the Ztgf β -9 polypeptides
described above; and (b) a nucleotide sequence
30 complementary to any of the nucleotide sequences in (a).

Further embodiments of the invention include
isolated nucleic acid molecules that comprise a
polynucleotide having a nucleotide sequence at least 90%
35 homologous, and more preferably 95%, 97%, 98%, or 99%
homologous to any of the nucleotide sequences in (a) or
(b) above, or a polynucleotide which hybridizes under

stringent hybridization conditions to a polynucleotide having a nucleotide sequence of (a) or (b) above.

Further embodiments of the invention include
5 isolated polypeptides having an amino acid sequence that is at least 90% identical, and more preferably 95%, 97%, 98%, or 99% identical to any of the Ztgf β -9 polypeptides and polynucleotides which encode these polypeptides.

10 Within another aspect of the invention there is provided an expression vector comprising (a) a transcription promoter; (b) a DNA segment encoding a polypeptide described above, and (c) a transcription terminator, wherein the promoter, DNA segment, and
15 terminator are operably linked.

Within a third aspect of the invention there is provided a cultured eukaryotic cell into which has been introduced an expression vector as disclosed above,
20 wherein said cell expresses a protein polypeptide encoded by the DNA segment.

In another embodiment of the present invention is an isolated antibody that binds specifically to a
25 Ztgf β -9 polypeptide described above. Also claimed is a method for producing antibodies which bind to a Ztgf β -9 polypeptide comprising inoculating a mammal with a Ztgf β -9 polypeptide or Ztgf β -9 epitope-bearing polypeptide so that the mammal produces antibodies to the polypeptide; and
30 isolating said antibodies.

These and other aspects of the invention will become evident upon reference to the following detailed description.

DETAILED DESCRIPTION OF THE INVENTION

The teachings of all of the references cited herein are incorporated in their entirety herein by
5 reference.

In the description that follows, a number of terms are used extensively. The following definitions are provided to facilitate understanding of the invention.

10

As used herein, "nucleic acid" or "nucleic acid molecule" refers to polynucleotides, such as deoxyribonucleic acid (DNA) or ribonucleic acid (RNA), oligonucleotides, fragments generated by the polymerase
15 chain reaction (PCR), and fragments generated by any of ligation, scission, endonuclease action, and exonuclease action. Nucleic acid molecules can be composed of monomers that are naturally-occurring nucleotides (such as DNA and RNA), or analogs of naturally-occurring
20 nucleotides (e.g., α -enantiomeric forms of naturally-occurring nucleotides), or a combination of both. Modified nucleotides can have alterations in sugar moieties and/or in pyrimidine or purine base moieties. Sugar modifications include, for example, replacement of
25 one or more hydroxyl groups with halogens, alkyl groups, amines, and azido groups, or sugars can be functionalized as ethers or esters. Moreover, the entire sugar moiety can be replaced with sterically and electronically similar structures, such as aza-sugars and carbocyclic sugar
30 analogs. Examples of modifications in a base moiety include alkylated purines and pyrimidines, acylated purines or pyrimidines, or other well-known heterocyclic substitutes. Nucleic acid monomers can be linked by phosphodiester bonds or analogs of such linkages. Analogs
35 of phosphodiester linkages include phosphorothioate, phosphorodithioate, phosphoroselenoate, phosphorodiselenoate, phosphoroanilothioate,

phosphoranilidate, phosphoramidate, and the like. The term "nucleic acid molecule" also includes so-called "peptide nucleic acids," which comprise naturally-occurring or modified nucleic acid bases attached to a polyamide backbone. Nucleic acids can be either single stranded or double stranded.

The term "complement of a nucleic acid molecule" refers to a nucleic acid molecule having a complementary nucleotide sequence and reverse orientation as compared to a reference nucleotide sequence. For example, the sequence 5' ATGCACGGG 3' is complementary to 5' CCCGTGCAT 3'.

The term "contig" denotes a nucleic acid molecule that has a contiguous stretch of identical or complementary sequence to another nucleic acid molecule. Contiguous sequences are said to "overlap" a given stretch of a nucleic acid molecule either in their entirety or along a partial stretch of the nucleic acid molecule.

The term "degenerate nucleotide sequence" denotes a sequence of nucleotides that includes one or more degenerate codons as compared to a reference nucleic acid molecule that encodes a polypeptide. Degenerate codons contain different triplets of nucleotides, but encode the same amino acid residue (*i.e.*, GAU and GAC triplets each encode Asp).

The term "structural gene" refers to a nucleic acid molecule that is transcribed into messenger RNA (mRNA), which is then translated into a sequence of amino acids characteristic of a specific polypeptide.

An "isolated nucleic acid molecule" is a nucleic acid molecule that is not integrated in the genomic DNA of an organism. For example, a DNA molecule that encodes a

growth factor that has been separated from the genomic DNA of a cell is an isolated DNA molecule. Another example of an isolated nucleic acid molecule is a chemically-synthesized nucleic acid molecule that is not integrated in the genome of an organism. A nucleic acid molecule that has been isolated from a particular species is smaller than the complete DNA molecule of a chromosome from that species.

10 A "nucleic acid molecule construct" is a nucleic acid molecule, either single- or double-stranded, that has been modified through human intervention to contain segments of nucleic acid combined and juxtaposed in an arrangement not existing in nature.

15 "Linear DNA" denotes non-circular DNA molecules having free 5' and 3' ends. Linear DNA can be prepared from closed circular DNA molecules, such as plasmids, by enzymatic digestion or physical disruption.

20 "Complementary DNA (cDNA)" is a single-stranded DNA molecule that is formed from an mRNA template by the enzyme reverse transcriptase. Typically, a primer complementary to portions of mRNA is employed for the initiation of reverse transcription. Those skilled in the art also use the term "cDNA" to refer to a double-stranded DNA molecule consisting of such a single-stranded DNA molecule and its complementary DNA strand. The term "cDNA" also refers to a clone of a cDNA molecule synthesized from an RNA template.

35 A "promoter" is a nucleotide sequence that directs the transcription of a structural gene. Typically, a promoter is located in the 5' non-coding region of a gene, proximal to the transcriptional start site of a structural gene. Sequence elements within promoters that function in the initiation of transcription are often

characterized by consensus nucleotide sequences. These promoter elements include RNA polymerase binding sites, TATA sequences, CAAT sequences, differentiation-specific elements (DSEs; McGehee et al., *Mol. Endocrinol.* 7:551
 5 (1993)), cyclic AMP response elements (CREs), serum response elements (SREs; Treisman, *Seminars in Cancer Biol.* 1:47 (1990)), glucocorticoid response elements (GRES), and binding sites for other transcription factors, such as CRE/ATF (O'Reilly et al., *J. Biol. Chem.* 267:19938
 10 (1992)), AP2 (Ye et al., *J. Biol. Chem.* 269:25728 (1994)), SP1, cAMP response element binding protein (CREB; Loeken, *Gene Expr.* 3:253 (1993)) and octamer factors (see, in general, Watson et al., eds., *Molecular Biology of the Gene*, 4th ed. (The Benjamin/Cummings Publishing Company,
 15 Inc. 1987), and Lemaigre and Rousseau, *Biochem. J.* 303:1 (1994)). If a promoter is an inducible promoter, then the rate of transcription increases in response to an inducing agent. In contrast, the rate of transcription is not regulated by an inducing agent if the promoter is a
 20 constitutive promoter. Repressible promoters are also known.

A "core promoter" contains essential nucleotide sequences for promoter function, including the TATA box
 25 and start of transcription. By this definition, a core promoter may or may not have detectable activity in the absence of specific sequences that may enhance the activity or confer tissue specific activity.

30 A "regulatory element" is a nucleotide sequence that modulates the activity of a core promoter. For example, a regulatory element may contain a nucleotide sequence that binds with cellular factors enabling transcription exclusively or preferentially in particular
 35 cells, tissues, or organelles. These types of regulatory elements are normally associated with genes that are expressed in a "cell-specific," "tissue-specific," or

"organelle-specific" manner. For example, the Ztgf β -9 regulatory element preferentially induces gene expression in brain, spinal cord, heart, skeletal muscle, stomach, pancreas, adrenal gland, salivary gland, liver, small intestine, bone marrow, thymus, spleen, lymph node, heart, thyroid, trachea, testis, ovary and placenta.

An "enhancer" is a type of regulatory element that can increase the efficiency of transcription, regardless of the distance or orientation of the enhancer relative to the start site of transcription.

"Heterologous DNA" refers to a DNA molecule, or a population of DNA molecules, that does not exist naturally within a given host cell. DNA molecules heterologous to a particular host cell may contain DNA derived from the host cell species (*i.e.*, endogenous DNA) so long as that host DNA is combined with non-host DNA (*i.e.*, exogenous DNA). For example, a DNA molecule containing a non-host DNA segment encoding a polypeptide operably linked to a host DNA segment comprising a transcription promoter is considered to be a heterologous DNA molecule. Conversely, a heterologous DNA molecule can comprise an endogenous gene operably linked with an exogenous promoter. As another illustration, a DNA molecule comprising a gene derived from a wild-type cell is considered to be heterologous DNA if that DNA molecule is introduced into a mutant cell that lacks the wild-type gene.

30

A "polypeptide" is a polymer of amino acid residues joined by peptide bonds, whether produced naturally or synthetically. Polypeptides of less than about 10 amino acid residues are commonly referred to as "peptides."

35

A "protein" is a macromolecule comprising one or more polypeptide chains. A protein may also comprise non-peptidic components, such as carbohydrate groups.

Carbohydrates and other non-peptidic substituents may be added to a protein by the cell in which the protein is produced, and will vary with the type of cell. Proteins are defined herein in terms of their amino acid backbone structures; substituents such as carbohydrate groups are generally not specified, but may be present nonetheless.

A peptide or polypeptide encoded by a non-host DNA molecule is a "heterologous" peptide or polypeptide.

An "integrated genetic element" is a segment of DNA that has been incorporated into a chromosome of a host cell after that element is introduced into the cell through human manipulation. Within the present invention, integrated genetic elements are most commonly derived from linearized plasmids that are introduced into the cells by electroporation or other techniques. Integrated genetic elements are passed from the original host cell to its progeny.

A "cloning vector" is a nucleic acid molecule, such as a plasmid, cosmid, or bacteriophage, that has the capability of replicating autonomously in a host cell. Cloning vectors typically contain one or a small number of restriction endonuclease recognition sites that allow insertion of a nucleic acid molecule in a determinable fashion without loss of an essential biological function of the vector, as well as nucleotide sequences encoding a marker gene that is suitable for use in the identification and selection of cells transformed with the cloning vector. Marker genes typically include genes that provide tetracycline resistance or ampicillin resistance.

An "expression vector" is a nucleic acid molecule encoding a gene that is expressed in a host cell.

Typically, an expression vector comprises a transcription promoter, a gene, and a transcription terminator. Gene
5 expression is usually placed under the control of a promoter, and such a gene is said to be "operably linked to" the promoter. Similarly, a regulatory element and a core promoter are operably linked if the regulatory element modulates the activity of the core promoter.

10

A "recombinant host" is a cell that contains a heterologous nucleic acid molecule, such as a cloning vector or expression vector. In the present context, an
15 example of a recombinant host is a cell that produces Ztgf β -9 from an expression vector. In contrast, Ztgf β -9 can be produced by a cell that is a "natural source" of Ztgf β -9, and that lacks an expression vector.

"Integrative transformants" are recombinant host
20 cells, in which heterologous DNA has become integrated into the genomic DNA of the cells.

A "fusion protein" is a hybrid protein expressed by a nucleic acid molecule comprising nucleotide sequences
25 of at least two genes. For example, a fusion protein can comprise at least part of a Ztgf β -9 polypeptide fused with a polypeptide that binds an affinity matrix. Such a fusion protein provides a means to isolate large quantities of Ztgf β -9 using affinity chromatography.

30

The term "receptor" denotes a cell-associated protein that binds to a bioactive molecule termed a "ligand." This interaction mediates the effect of the ligand on the cell. Receptors can be membrane bound,
35 cytosolic or nuclear; monomeric (e.g., thyroid stimulating hormone receptor, beta-adrenergic receptor) or multimeric (e.g., PDGF receptor, growth hormone receptor, IL-3

receptor, GM-CSF receptor, G-CSF receptor, erythropoietin receptor and IL-6 receptor). Membrane-bound receptors are characterized by a multi-domain structure comprising an extracellular ligand-binding domain and an intracellular effector domain that is typically involved in signal transduction. In certain membrane-bound receptors, the extracellular ligand-binding domain and the intracellular effector domain are located in separate polypeptides that comprise the complete functional receptor.

In general, the binding of ligand to receptor results in a conformational change in the receptor that causes an interaction between the effector domain and other molecule(s) in the cell, which in turn leads to an alteration in the metabolism of the cell. Metabolic events that are often linked to receptor-ligand interactions include gene transcription, phosphorylation, dephosphorylation, increases in cyclic AMP production, mobilization of cellular calcium, mobilization of membrane lipids, cell adhesion, hydrolysis of inositol lipids and hydrolysis of phospholipids.

The term "secretory signal sequence" denotes a DNA sequence that encodes a peptide (a "secretory peptide") that, as a component of a larger polypeptide, directs the larger polypeptide through a secretory pathway of a cell in which it is synthesized. The larger polypeptide is commonly cleaved to remove the secretory peptide during transit through the secretory pathway.

An "isolated polypeptide" is a polypeptide that is essentially free from contaminating cellular components, such as carbohydrate, lipid, or other proteinaceous impurities associated with the polypeptide in nature. Typically, a preparation of isolated polypeptide contains the polypeptide in a highly purified form, i.e., at least about 80% pure, at least about 90%

pure, at least about 95% pure, greater than 95% pure, or greater than 99% pure. One way to show that a particular protein preparation contains an isolated polypeptide is by the appearance of a single band following sodium dodecyl sulfate (SDS)-polyacrylamide gel electrophoresis of the protein preparation and Coomassie Brilliant Blue staining of the gel. However, the term "isolated" does not exclude the presence of the same polypeptide in alternative physical forms, such as dimers or alternatively glycosylated or derivatized forms.

The terms "amino-terminal or N-terminal" and "carboxyl-terminal or C-terminal" are used herein to denote positions within polypeptides. Where the context allows, these terms are used with reference to a particular sequence or portion of a polypeptide to denote proximity or relative position. For example, a certain sequence positioned carboxyl-terminal to a reference sequence within a polypeptide is located proximal to the carboxyl terminus of the reference sequence, but is not necessarily at the carboxyl terminus of the complete polypeptide.

The term "expression" refers to the biosynthesis of a gene product. For example, in the case of a structural gene, expression involves transcription of the structural gene into mRNA and the translation of mRNA into one or more polypeptides.

The term "splice variant" is used herein to denote alternative forms of RNA transcribed from a gene. Splice variation arises naturally through use of alternative splicing sites within a transcribed RNA molecule, or less commonly between separately transcribed RNA molecules, and may result in several mRNAs transcribed from the same gene. Splice variants may encode polypeptides having altered amino acid sequence. The term

splice variant is also used herein to denote a polypeptide encoded by a splice variant of an mRNA transcribed from a gene.

5 As used herein, the term "immunomodulator" includes cytokines, stem cell growth factors, lymphotoxins, co-stimulatory molecules, hematopoietic factors, and synthetic analogs of these molecules.

10 The term "complement/anti-complement pair" denotes non-identical moieties that form a non-covalently associated, stable pair under appropriate conditions. For instance, biotin and avidin (or streptavidin) are prototypical members of a complement/anti-complement pair.
 15 Other exemplary complement/anti-complement pairs include receptor/ligand pairs, antibody/antigen (or hapten or epitope) pairs, sense/antisense polynucleotide pairs, and the like. Where subsequent dissociation of the complement/anti-complement pair is desirable, the
 20 complement/anti-complement pair preferably has a binding affinity of less than 10^9 M^{-1} .

 An "anti-idiotypic antibody" is an antibody that binds with the variable region domain of an
 25 immunoglobulin. In the present context, an anti-idiotypic antibody binds with the variable region of an anti- Ztgf β -9 antibody, and thus, an anti-idiotypic antibody mimics an epitope of Ztgf β -9.

30 An "antibody fragment" is a portion of an antibody such as F(ab')_2 , F(ab)_2 , Fab' , Fab , and the like. Regardless of structure, an antibody fragment binds with the same antigen that is recognized by the intact antibody. For example, a Ztgf β -9 monoclonal antibody fragment binds
 35 with an epitope of Ztgf β -9.

The term "antibody fragment" also includes a synthetic or a genetically engineered polypeptide that binds to a specific antigen, such as polypeptides consisting of the light chain variable region, "Fv" fragments consisting of the variable regions of the heavy and light chains, recombinant single chain polypeptide molecules in which light and heavy variable regions are connected by a peptide linker ("scFv proteins"), and minimal recognition units consisting of the amino acid residues that mimic the hypervariable region.

A "chimeric antibody" is a recombinant protein that contains the variable domains and complementary determining regions derived from a rodent antibody, while the remainder of the antibody molecule is derived from a human antibody.

"Humanized antibodies" are recombinant proteins in which murine complementarity determining regions of a monoclonal antibody have been transferred from heavy and light variable chains of the murine immunoglobulin into a human variable domain.

As used herein, a "therapeutic agent" is a molecule or atom which is conjugated to an antibody moiety to produce a conjugate which is useful for therapy. Examples of therapeutic agents include drugs, toxins, immunomodulators, chelators, boron compounds, photoactive agents or dyes, and radioisotopes.

A "detectable label" is a molecule or atom which can be conjugated to an antibody moiety to produce a molecule useful for diagnosis. Examples of detectable labels include chelators, photoactive agents, radioisotopes, fluorescent agents, paramagnetic ions, or other marker moieties.

The term "affinity tag" is used herein to denote a polypeptide segment that can be attached to a second polypeptide to provide for purification or detection of the second polypeptide or provide sites for attachment of the second polypeptide to a substrate. In principal, any peptide or protein for which an antibody or other specific binding agent is available can be used as an affinity tag. Affinity tags include a poly-histidine tract, protein A (Nilsson et al., *EMBO J.* 4:1075 (1985); Nilsson et al., *Methods Enzymol.* 198:3 (1991)), glutathione S transferase (Smith and Johnson, *Gene* 67:31 (1988)), Glu-Glu affinity tag (Grussenmeyer et al., *Proc. Natl. Acad. Sci. USA* 82:7952 (1985)), substance P, FLAG peptide (Hopp et al., *Biotechnology* 6:1204 (1988)), streptavidin binding peptide, or other antigenic epitope or binding domain. See, in general, Ford et al., *Protein Expression and Purification* 2:95 (1991). DNAs encoding affinity tags are available from commercial suppliers (e.g., Pharmacia Biotech, Piscataway, NJ).

20

A "naked antibody" is an entire antibody, as opposed to an antibody fragment, which is not conjugated with a therapeutic agent. Naked antibodies include both polyclonal and monoclonal antibodies, as well as certain recombinant antibodies, such as chimeric and humanized antibodies.

25

As used herein, the term "antibody component" includes both an entire antibody and an antibody fragment.

30

An "immunoconjugate" is a conjugate of an antibody component with a therapeutic agent or a detectable label.

35

As used herein, the term "antibody fusion protein" refers to a recombinant molecule that comprises an antibody component and a therapeutic agent. Examples

of therapeutic agents suitable for such fusion proteins include immunomodulators ("antibody-immunomodulator fusion protein") and toxins ("antibody-toxin fusion protein").

5 A "tumor associated antigen" is a protein normally not expressed, or expressed at lower levels, by a normal counterpart cell. Examples of tumor associated antigens include alpha-fetoprotein, carcinoembryonic antigen, and Her-2/neu. Many other illustrations of tumor
10 associated antigens are known to those of skill in the art. See, for example, Urban et al., *Ann. Rev. Immunol.* 10:617 (1992).

 As used herein, an "infectious agent" denotes
15 both microbes and parasites. A "microbe" includes viruses, bacteria, rickettsia, mycoplasma, protozoa, fungi and like microorganisms. A "parasite" denotes infectious, generally microscopic or very small multicellular invertebrates, or ova or juvenile forms thereof, which are
20 susceptible to immune-mediated clearance or lytic or phagocytic destruction, such as malarial parasites, spirochetes, and the like.

 An "infectious agent antigen" is an antigen
25 associated with an infectious agent.

 A "target polypeptide" or a "target peptide" is an amino acid sequence that comprises at least one epitope, and that is expressed on a target cell, such as a
30 tumor cell, or a cell that carries an infectious agent antigen. T-cells recognize peptide epitopes presented by a major histocompatibility complex molecule to a target polypeptide or target peptide and typically lyse the target cell or recruit other immune cells to the site of
35 the target cell, thereby killing the target cell.

An "antigenic peptide" is a peptide which will bind a major histocompatibility complex molecule to form an MHC-peptide complex which is recognized by a T cell, thereby inducing a cytotoxic lymphocyte response upon presentation to the T cell. Thus, antigenic peptides are capable of binding to an appropriate major histocompatibility complex molecule and inducing a cytotoxic T cells response, such as cell lysis or specific cytokine release against the target cell which binds or expresses the antigen. The antigenic peptide can be bound in the context of a class I or class II major histocompatibility complex molecule, on an antigen presenting cell or on a target cell.

In eukaryotes, RNA polymerase II catalyzes the transcription of a structural gene to produce mRNA. A nucleic acid molecule can be designed to contain an RNA polymerase II template in which the RNA transcript has a sequence that is complementary to that of a specific mRNA. The RNA transcript is termed an "anti-sense RNA" and a nucleic acid molecule that encodes the anti-sense RNA is termed an "anti-sense gene." Anti-sense RNA molecules are capable of binding to mRNA molecules, resulting in an inhibition of mRNA translation or mRNA degradation.

An "anti-sense oligonucleotide specific for *Ztgf β -9*" or a "*Ztgf β -9* anti-sense oligonucleotide" is an oligonucleotide having a sequence (a) capable of forming a stable triplex with a portion of the *Ztgf β -9* gene, or (b) capable of forming a stable duplex with a portion of an mRNA transcript of the *Ztgf β -9* gene.

A "ribozyme" is a nucleic acid molecule that contains a catalytic center. The term includes RNA enzymes, self-splicing RNAs, self-cleaving RNAs, and nucleic acid molecules that perform these catalytic

functions. A nucleic acid molecule that encodes a ribozyme is termed a "ribozyme gene."

5 An "external guide sequence" is a nucleic acid molecule that directs the endogenous ribozyme, RNase P, to a particular species of intracellular mRNA, resulting in the cleavage of the mRNA by RNase P. A nucleic acid molecule that encodes an external guide sequence is termed an "external guide sequence gene."

10

The term "variant human *Ztgfb-9* gene" refers to nucleic acid molecules that encode a polypeptide having an amino acid sequence that is a modification of SEQ ID NO:2. Such variants include naturally-occurring polymorphisms of
15 *Ztgfb-9* genes, as well as synthetic genes that contain conservative amino acid substitutions of the amino acid sequence of SEQ ID NO:2. Additional variant forms of *Ztgfb-9* genes are nucleic acid molecules that contain insertions or deletions of the nucleotide sequences
20 described herein. A variant *Ztgfb-9* gene can be identified by determining whether the gene hybridizes with a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1, or its complement, under stringent conditions.

25

Similarly, the term "variant murine *Ztgfb-9* gene" refers to nucleic acid molecules that encode a polypeptide having an amino acid sequence that is a modification of SEQ ID NO:9. A variant murine *Ztgfb-9*
30 gene can be identified by determining whether the gene hybridizes with a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:8, or its complement, under stringent conditions.

35

Alternatively, variant *Ztgfb-9* genes can be identified by sequence comparison. Two amino acid sequences have "100% amino acid sequence identity" if the

amino acid residues of the two amino acid sequences are the same when aligned for maximal correspondence. Similarly, two nucleotide sequences have "100% nucleotide sequence identity" if the nucleotide residues of the two

5 nucleotide sequences are the same when aligned for maximal correspondence. Sequence comparisons can be performed using standard software programs such as those included in the LASERGENE bioinformatics computing suite, which is produced by DNASTAR (Madison, Wisconsin). Other methods

10 for comparing two nucleotide or amino acid sequences by determining optimal alignment are well-known to those of skill in the art (see, for example, Peruski and Peruski, *The Internet and the New Biology: Tools for Genomic and Molecular Research* (ASM Press, Inc. 1997), Wu et al.

15 (eds.), "Information Superhighway and Computer Databases of Nucleic Acids and Proteins," in *Methods in Gene Biotechnology*, pages 123-151 (CRC Press, Inc. 1997), and Bishop (ed.), *Guide to Human Genome Computing*, 2nd Edition (Academic Press, Inc. 1998)). Particular methods for

20 determining sequence identity are described below.

Regardless of the particular method used to identify a variant *Ztgf β -9* gene or variant *Ztgf β -9* polypeptide, a variant gene or polypeptide encoded by a

25 variant gene is functionally characterized by either its anti-viral or anti-proliferative activities, or by the ability to bind specifically to an anti-*Ztgf β -9* antibody.

The term "allelic variant" is used herein to

30 denote any of two or more alternative forms of a gene occupying the same chromosomal locus. Allelic variation arises naturally through mutation, and may result in phenotypic polymorphism within populations. Gene mutations can be silent (no change in the encoded

35 polypeptide) or may encode polypeptides having altered amino acid sequence. The term allelic variant is also used

herein to denote a protein encoded by an allelic variant of a gene.

5 The term "ortholog" denotes a polypeptide or protein obtained from one species that is the functional counterpart of a polypeptide or protein from a different species. Sequence differences among orthologs are the result of speciation.

10 "Paralogs" are distinct but structurally related proteins made by an organism. Paralogs are believed to arise through gene duplication. For example, α -globin, β -globin, and myoglobin are paralogs of each other.

15 Due to the imprecision of standard analytical methods, molecular weights and lengths of polymers are understood to be approximate values. When such a value is expressed as "about" X or "approximately" X, the stated value of X will be understood to be accurate to $\pm 10\%$.

20 Nucleic acid molecules encoding a human or mouse *Ztgf β -9* gene can be obtained by screening a human or mouse cDNA or genomic library using polynucleotide probes based upon SEQ ID NO:1 or SEQ ID NO:8. These techniques are
25 standard and well-established. As an illustration, a nucleic acid molecule that encodes a human *Ztgf β -9* gene can be isolated from a human cDNA library. In this case, the first step would be to prepare the cDNA library by isolating RNA from brain, spinal cord, heart, skeletal
30 muscle, stomach, pancreas, adrenal gland, salivary gland, liver, small intestine, bone marrow, thymus, spleen, lymph node, heart, thyroid, trachea, testis, ovary or placental tissue, using methods well-known to those of skill in the art. In general, RNA isolation techniques must provide a
35 method for breaking cells, a means of inhibiting RNase-directed degradation of RNA, and a method of separating RNA from DNA, protein, and polysaccharide contaminants. For

example, total RNA can be isolated by freezing tissue in liquid nitrogen, grinding the frozen tissue with a mortar and pestle to lyse the cells, extracting the ground tissue with a solution of phenol/chloroform to remove proteins, and separating RNA from the remaining impurities by selective precipitation with lithium chloride (see, for example, Ausubel et al. (eds.), *Short Protocols in Molecular Biology*, 3rd Edition, pages 4-1 to 4-6 (John Wiley & Sons 1995) ["Ausubel (1995)"]; Wu et al., *Methods in Gene Biotechnology*, pages 33-41 (CRC Press, Inc. 1997) ["Wu (1997)"]).

Alternatively, total RNA can be isolated from brain or spinal cord tissue as well as heart, skeletal muscle, stomach, pancreas, adrenal gland, salivary gland, liver, small intestine, bone marrow, thymus, spleen, lymph node, thyroid, trachea, testis, ovary or placenta by extracting ground tissue with guanidinium isothiocyanate, extracting with organic solvents, and separating RNA from contaminants using differential centrifugation (see, for example, Chirgwin et al., *Biochemistry* 18:52 (1979); Ausubel (1995) at pages 4-1 to 4-6; Wu (1997) at pages 33-41). To construct a cDNA library, poly(A)⁺ RNA must be isolated from a total RNA preparation. Poly(A)⁺ RNA can be isolated from total RNA using the standard technique of oligo(dT)-cellulose chromatography (see, for example, Aviv and Leder, *Proc. Nat'l Acad. Sci. USA* 69:1408 (1972); Ausubel (1995) at pages 4-11 to 4-12). Double-stranded cDNA molecules are synthesized from poly(A)⁺ RNA using techniques well-known to those in the art. (see, for example, Wu (1997) at pages 41-46). Moreover, commercially available kits can be used to synthesize double-stranded cDNA molecules. For example, such kits are available from Life Technologies, Inc. (Gaithersburg, MD), CLONTECH Laboratories, Inc. (Palo Alto, CA), Promega Corporation (Madison, WI) and STRATAGENE (La Jolla, CA).

Various cloning vectors are appropriate for the construction of a cDNA library. For example, a cDNA library can be prepared in a vector derived from bacteriophage, such as a λ gt10 vector. See, for example, 5 Huynh et al., "Constructing and Screening cDNA Libraries in λ gt10 and λ gt11," in *DNA Cloning: A Practical Approach Vol. I*, Glover (ed.), page 49 (IRL Press, 1985); Wu (1997) at pages 47-52. Alternatively, double-stranded cDNA molecules can be inserted into a plasmid vector, such as a 10 pBLUESCRIPT vector (STRATAGENE; La Jolla, CA), a LAMDAGEM-4 (Promega Corp.) or other commercially available vectors. Suitable cloning vectors also can be obtained from the American Type Culture Collection (Manassas, VA). To amplify the cloned cDNA molecules, the cDNA library is inserted 15 into a prokaryotic host, using standard techniques. For example, a cDNA library can be introduced into competent *E. coli* DH5 cells, which can be obtained, for example, from Life Technologies, Inc. (Gaithersburg, MD).

20 A human genomic library can be prepared by means well-known in the art (see, for example, Ausubel (1995) at pages 5-1 to 5-6; Wu (1997) at pages 307-327). Genomic DNA can be isolated by lysing tissue with the detergent Sarkosyl, digesting the lysate with proteinase K, clearing 25 insoluble debris from the lysate by centrifugation, precipitating nucleic acid from the lysate using isopropanol, and purifying resuspended DNA on a cesium chloride density gradient. DNA fragments that are suitable for the production of a genomic library can be obtained by 30 the random shearing of genomic DNA or by the partial digestion of genomic DNA with restriction endonucleases. Genomic DNA fragments can be inserted into a vector, such as a bacteriophage or cosmid vector, in accordance with conventional techniques, such as the use of restriction 35 enzyme digestion to provide appropriate termini, the use of alkaline phosphatase treatment to avoid undesirable joining of DNA molecules, and ligation with appropriate ligases.

Techniques for such manipulation are well-known in the art (see, for example, Ausubel (1995) at pages 5-1 to 5-6; Wu (1997) at pages 307-327).

5 Nucleic acid molecules that encode a human *Ztgf β -9* gene can also be obtained using the polymerase chain reaction (PCR) with oligonucleotide primers having nucleotide sequences that are based upon the nucleotide sequences of the human *Ztgf β -9* gene, as described herein.

10 General methods for screening libraries with PCR are provided by, for example, Yu et al., "Use of the Polymerase Chain Reaction to Screen Phage Libraries," in *Methods in Molecular Biology, Vol. 15: PCR Protocols: Current Methods and Applications*, White (ed.), pages 211-

15 215 (Humana Press, Inc. 1993). Moreover, techniques for using PCR to isolate related genes are described by, for example, Preston, "Use of Degenerate Oligonucleotide Primers and the Polymerase Chain Reaction to Clone Gene Family Members," in *Methods in Molecular Biology, Vol. 15: PCR Protocols: Current Methods and Applications*, White

20 (ed.), pages 317-337 (Humana Press, Inc. 1993). Alternatively, human genomic libraries can be obtained from commercial sources such as Research Genetics (Huntsville, AL) and the American Type Culture Collection (Manassas,

25 VA). A library containing cDNA or genomic clones can be screened with one or more polynucleotide probes based upon SEQ ID NO:1, using standard methods (see, for example, Ausubel (1995) at pages 6-1 to 6-11).

30 Anti- *Ztgf β -9* antibodies, produced as described below, can also be used to isolate DNA sequences that encode human *Ztgf β -9* genes from cDNA libraries. For example, the antibodies can be used to screen λ gt11 expression libraries, or the antibodies can be used for

35 immunoscreening following hybrid selection and translation (see, for example, Ausubel (1995) at pages 6-12 to 6-16; Margolis et al., "Screening λ expression libraries with

antibody and protein probes," in *DNA Cloning 2: Expression Systems, 2nd Edition*, Glover et al. (eds.), pages 1-14 (Oxford University Press 1995)).

5 As an alternative, an *Ztgf β -9* gene can be obtained by synthesizing nucleic acid molecules using mutually priming long oligonucleotides and the nucleotide sequences described herein (see, for example, Ausubel (1995) at pages 8-8 to 8-9). Established techniques using
10 the polymerase chain reaction provide the ability to synthesize DNA molecules at least two kilobases in length (Adang et al., *Plant Molec. Biol.* 21:1131 (1993), Bambot et al., *PCR Methods and Applications* 2:266 (1993), Dillon et al., "Use of the Polymerase Chain Reaction for the
15 Rapid Construction of Synthetic Genes," in *Methods in Molecular Biology, Vol. 15: PCR Protocols: Current Methods and Applications*, White (ed.), pages 263-268, (Humana Press, Inc. 1993), and Holowachuk et al., *PCR Methods Appl.* 4:299 (1995)). The sequence of an *Ztgf β -9* cDNA or
20 *Ztgf β -9* genomic fragment can be determined using standard methods. Moreover, the identification of genomic fragments containing an *Ztgf β -9* promoter or regulatory element can be achieved using well-established techniques, such as deletion analysis (see, generally, Ausubel
25 (1995)).

Cloning of 5' flanking sequences also facilitates production of *Ztgf β -9* proteins by "gene activation," following the methods disclosed in U.S.
30 Patent No. 5,641,670. Briefly, expression of an endogenous *Ztgf β -9* gene in a cell is altered by introducing into the *Ztgf β -9* locus a DNA construct comprising at least a targeting sequence, a regulatory sequence, an exon, and an unpaired splice donor site. The targeting sequence is
35 a *Ztgf β -9* 5' non-coding sequence that permits homologous recombination of the construct with the endogenous *Ztgf β -9* locus, whereby the sequences within the construct become

operably linked with the endogenous *Ztgf β -9* coding sequence. In this way, an endogenous *Ztgf β -9* promoter can be replaced or supplemented with other regulatory sequences to provide enhanced, tissue-specific, or
 5 otherwise regulated expression.

Additionally, the polynucleotides of the present invention can be synthesized using a DNA synthesizer. Currently the method of choice is the phosphoramidite
 10 method. If chemically synthesized double stranded DNA is required for an application such as the synthesis of a gene or a gene fragment, then each complementary strand is made separately. The production of short genes (60 to 80 bp) is technically straightforward and can be accomplished
 15 by synthesizing the complementary strands and then annealing them. For the production of longer genes (>300 bp), however, special strategies must be invoked, because the coupling efficiency of each cycle during chemical DNA synthesis is seldom 100%. To overcome this problem,
 20 synthetic genes (double-stranded) are assembled in modular form from single-stranded fragments that are from 20 to 100 nucleotides in length. In addition to the protein coding sequence, synthetic genes can be designed with terminal sequences that facilitate insertion into a
 25 restriction endonuclease sites of a cloning vector and other sequences should also be added that contain signals for the proper initiation and termination of transcription and translation. See Glick, Bernard R. and Jack J. Pasternak, *Molecular Biotechnology, Principles &*
 30 *Applications of Recombinant DNA*, (ASM Press, Washington, D.C. 1994), Itakura, K. et al. Synthesis and use of synthetic oligonucleotides. *Annu. Rev. Biochem.* 53 : 323-356 (1984), and Climie, S. et al. Chemical synthesis of the thymidylate synthase gene. *Proc. Natl. Acad. Sci. USA*
 35 87 :633-637 (1990).

Within preferred embodiments of the invention the isolated polynucleotides will hybridize to similar sized regions of the DNA of SEQ ID NO:1, or a sequence complementary thereto, under stringent conditions. In
5 general, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly
10 matched probe. Typical stringent conditions are those in which the salt concentration is about 0.02 M or less at pH 7 and the temperature is at least about 60°C. As previously noted, the isolated polynucleotides of the present invention include DNA and RNA. Methods for
15 isolating DNA and RNA are well known in the art. Total RNA can be prepared using guanidine HCl extraction followed by isolation by centrifugation in a CsCl gradient [Chirgwin *et al.*, *Biochemistry* 18:52-94 (1979)]. Poly (A)⁺ RNA is prepared from total RNA using the method of Aviv and
20 Leder, *Proc. Natl. Acad. Sci. USA* 69:1408-1412 (1972). Complementary DNA (cDNA) is prepared from poly(A)⁺ RNA using known methods. Polynucleotides encoding Ztgfβ-9 polypeptides are then identified and isolated by, for example, hybridization or PCR.

25

Those skilled in the art will recognize that the sequences disclosed in SEQ ID NOS:1 and 2 represent a single allele of the human. There are a number of naturally occurring mature N-terminal variants having the
30 leader sequence cleaved at differing positions. Allelic variants of these sequences can be cloned by probing cDNA or genomic libraries from different individuals according to standard procedures. The present invention further provides counterpart proteins and polynucleotides from
35 other species ("species orthologs"). Of particular interest are Ztgfβ-9 polypeptides from other mammalian species, including murine, porcine, ovine, bovine, canine,

feline, equine, and other primates. Species orthologs of the human Ztgfb β -9 protein can be cloned using information and compositions provided by the present invention in combination with conventional cloning techniques. For example, a cDNA can be cloned using mRNA obtained from a tissue or cell type that expresses the gene. Suitable sources of mRNA can be identified by probing Northern blots with probes designed from the sequences disclosed herein. A library is then prepared from mRNA of a positive tissue or cell line. A protein-encoding cDNA can then be isolated by a variety of methods, such as by probing with a complete or partial human cDNA or with one or more sets of degenerate probes based on the disclosed sequences. A cDNA can also be cloned using the polymerase chain reaction, or PCR (Mullis, U.S. Patent No. 4,683,202), using primers designed from the sequences disclosed herein. Within an additional method, the cDNA library can be used to transform or transfect host cells, and expression of the cDNA of interest can be detected with an antibody to the protein. Similar techniques can also be applied to the isolation of genomic clones. As used and claimed the language "an isolated polynucleotide which encodes a polypeptide, said polynucleotide being defined by SEQ ID NO: 2" includes all allelic variants and species orthologs of the polypeptide of SEQ ID NOs:2, 3, 4 and 5.

Within preferred embodiments of the invention, isolated nucleic acid molecules that encode human Ztgfb β -9 can hybridize to nucleic acid molecules having the nucleotide sequence of SEQ ID NO:1, or a sequence complementary thereto, under "stringent conditions." In general, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at

which 50% of the target sequence hybridizes to a perfectly matched probe.

As an illustration, a nucleic acid molecule
5 encoding a variant Ztgf β -9 polypeptide can be hybridized
with a nucleic acid molecule having the nucleotide
sequence of SEQ ID NO:1 (or its complement) at 42°C
overnight in a solution comprising 50% formamide, 5xSSC
(1xSSC: 0.15 M sodium chloride and 15 mM sodium citrate),
10 50 mM sodium phosphate (pH 7.6), 5x Denhardt's solution
(100x Denhardt's solution: 2% (w/v) Ficoll 400, 2% (w/v)
polyvinylpyrrolidone, and 2% (w/v) bovine serum albumin),
10% dextran sulfate, and 20 μ g/ml denatured, sheared
salmon sperm DNA. One of skill in the art can devise
15 variations of these hybridization conditions. For
example, the hybridization mixture can be incubated at a
higher temperature, such as about 65°C, in a solution that
does not contain formamide. Moreover, premixed
hybridization solutions are available (e.g., EXPRESSHYB
20 Hybridization Solution from CLONTECH Laboratories, Inc.),
and hybridization can be performed according to the
manufacturer's instructions. Following hybridization, the
nucleic acid molecules can be washed to remove non-
hybridized nucleic acid molecules under stringent
25 conditions, or under highly stringent conditions. Typical
stringent washing conditions include washing in a solution
of 0.5x - 2x SSC with 0.1% sodium dodecyl sulfate (SDS) at
55 - 65°C. That is, nucleic acid molecules encoding a
variant Ztgf β -9 polypeptide hybridize with a nucleic acid
30 molecule having the nucleotide sequence of SEQ ID NO:1 (or
its complement) under stringent washing conditions, in
which the wash stringency is equivalent to 0.5x - 2x SSC
with 0.1% SDS at 55 - 65°C, including 0.5x SSC with 0.1%
SDS at 55°C, or 2xSSC with 0.1% SDS at 65°C. One of skill
35 in the art can readily devise equivalent conditions, for
example, by substituting SSPE for SSC in the wash
solution.

Typical highly stringent washing conditions include washing in a solution of 0.1x - 0.2x SSC with 0.1% sodium dodecyl sulfate (SDS) at 50 - 65°C. In other words, nucleic acid molecules encoding a variant *Ztgfβ-9* polypeptide hybridize with a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1 (or its complement) under highly stringent washing conditions, in which the wash stringency is equivalent to 0.1x - 0.2x SSC with 0.1% SDS at 50 - 65°C, including 0.1x SSC with 0.1% SDS at 50°C, or 0.2xSSC with 0.1% SDS at 65°C.

The present invention also provides isolated *Ztgfβ-9* polypeptides that have a substantially similar sequence identity to the polypeptides of SEQ ID NOs:2,3, 4, 5, 9, 12, 17, 18 or their orthologs. The term "substantially similar sequence identity" is used herein to denote polypeptides having at least 70%, at least 80%, at least 90%, at least 95% or greater than 95% and 99% sequence identity to the sequences shown in SEQ ID NOs:2, 3, 4, 5, 9, 12, 17, 18 or their orthologs.

The present invention also contemplates *Ztgfβ-9* variant nucleic acid molecules that can be identified using two criteria: a determination of the similarity between the encoded polypeptide with the amino acid sequence of SEQ ID NOs:2, 3, 4, 5, 9, 12, 17 or 18, and a hybridization assay, as described above. Such *Ztgfβ-9* variants include nucleic acid molecules (1) that hybridize with a nucleic acid molecule having the nucleotide sequence of ~~SEQ ID NO:1~~, SEQ ID NO: 9 or SEQ ID NO:16 (or their complement) under stringent washing conditions, in which the wash stringency is equivalent to 0.5x - 2x SSC with 0.1% SDS at 55 - 65°C, and (2) that encode a polypeptide having at least 70%, at least 80%, at least 90%, at least 95% or greater than 95% sequence identity to the amino acid sequence of SEQ ID NOs:2, 3, 4, 5, 9, 12,

17 or 18. Alternatively, *Ztgf β -9* variants can be characterized as nucleic acid molecules (1) that hybridize with a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1 (or its complement) under highly
 5 stringent washing conditions, in which the wash stringency is equivalent to 0.1x - 0.2x SSC with 0.1% SDS at 50 - 65°C, and (2) that encode a polypeptide having at least 70%, at least 80%, at least 90%, at least 95% or greater than 95% or 99% sequence identity to the amino acid
 10 sequence of SEQ ID NOs:2, 3, 4, 5, 9, 12, 17 or 18.

The present invention also contemplates human *Ztgf β -9* variant nucleic acid molecules identified by at least one of hybridization analysis and sequence identity
 15 determination, with reference to SEQ ID NO:2. The present invention further includes murine *Ztgf β -9* variant nucleic acid molecules identified by at least one of hybridization analysis and sequence identity determination, with reference to SEQ ID NOs:8 and 9. For example, using the
 20 approach discussed above, murine *Ztgf β -9* variant nucleic acid molecules can be identified using at least one of three criteria: (1) hybridization with a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:8 (or its complement) under stringent washing conditions, in
 25 which the wash stringency is equivalent to 0.5x - 2x SSC with 0.1% SDS at 55 - 65°C, (2) hybridization with a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:8 (or its complement) under highly stringent washing conditions, in which the wash stringency is
 30 equivalent to 0.1x - 0.2x SSC with 0.1% SDS at 50 - 65°C, and (3) an amino acid percent identity that is at least 70%, at least 80%, at least 90%, at least 95% or greater than 95% sequence identity to the amino acid sequence of SEQ ID NO:12.

35

Percent sequence identity is determined by conventional methods. See, for example, Altschul et al.,

10

Table 1

36

	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
A	4																			
R	-1	5																		
N	-2	0	6																	
D	-2	-2	1	6																
C	0	-3	-3	-3	9															
Q	-1	1	0	0	-3	5														
E	-1	0	0	2	-4	2	5													
G	0	-2	0	-1	-3	-2	-2	6												
H	-2	0	1	-1	-3	0	0	-2	8											
I	-1	-3	-3	-3	-1	-3	-3	-4	-3	4										
L	-1	-2	-3	-4	-1	-2	-3	-4	-3	2	4									
K	-1	2	0	-1	-3	1	1	-2	-1	-3	-2	5								
M	-1	-1	-2	-3	-1	0	-2	-3	-2	1	2	-1	5							
F	-2	-3	-3	-3	-2	-3	-3	-3	-1	0	0	-3	0	6						
P	-1	-2	-2	-1	-3	-1	-1	-2	-2	-3	-3	-1	-2	-4	7					
S	1	-1	1	0	-1	0	0	0	-1	-2	-2	0	-1	-2	-1	4				
T	0	-1	0	-1	-1	-1	-1	-2	-2	-1	-1	-1	-1	-2	-1	1	5			
W	-3	-3	-4	-4	-2	-2	-3	-2	-2	-3	-2	-3	-1	1	-4	-3	-2	11		
Y	-2	-2	-2	-3	-2	-1	-2	-3	2	-1	-1	-2	-1	3	-3	-2	-2	2	7	
V	0	-3	-3	-3	-1	-2	-2	-3	-3	3	1	-2	1	-1	-2	-2	0	-3	-1	4

5

10

15

20

Those skilled in the art appreciate that there are many established algorithms available to align two amino acid sequences. The "FASTA" similarity search algorithm of Pearson and Lipman is a suitable protein alignment method for examining the level of identity shared by an amino acid sequence disclosed herein and the amino acid sequence of a putative Ztgf β -9 variant. The FASTA algorithm is described by Pearson and Lipman, *Proc. Nat'l Acad. Sci. USA* 85:2444 (1988), and by Pearson, *Meth. Enzymol.* 183:63 (1990). Briefly, FASTA first characterizes sequence similarity by identifying regions shared by the query sequence (e.g., SEQ ID NO:2) and a test sequence that have either the highest density of identities (if the ktup variable is 1) or pairs of identities (if ktup=2), without considering conservative amino acid substitutions, insertions, or deletions. The ten regions with the highest density of identities are then re-scored by comparing the similarity of all paired amino acids using an amino acid substitution matrix, and the ends of the regions are "trimmed" to include only those residues that contribute to the highest score. If there are several regions with scores greater than the "cutoff" value (calculated by a predetermined formula based upon the length of the sequence and the ktup value), then the trimmed initial regions are examined to determine whether the regions can be joined to form an approximate alignment with gaps. Finally, the highest scoring regions of the two amino acid sequences are aligned using a modification of the Needleman-Wunsch-Sellers algorithm (Needleman and Wunsch, *J. Mol. Biol.* 48:444 (1970); Sellers, *SIAM J. Appl. Math.* 26:787 (1974)), which allows for amino acid insertions and deletions. Illustrative parameters for FASTA analysis are: ktup=1, gap opening penalty=10, gap extension penalty=1, and substitution matrix=BLOSUM62. These parameters can be introduced into a FASTA program by modifying the scoring matrix file

("SMATRIX"), as explained in Appendix 2 of Pearson, *Meth. Enzymol.* 183:63 (1990).

FASTA can also be used to determine the sequence
 5 identity of nucleic acid molecules using a ratio as
 disclosed above. For nucleotide sequence comparisons, the
 ktup value can range between one to six, preferably from
 three to six, most preferably three, with other parameters
 set as described above.

10

The present invention includes nucleic acid
 molecules that encode a polypeptide having a conservative
 amino acid change, compared with the amino acid sequence
 of SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO: 4, SEQ ID NO:5,
 15 SEQ ID NO:9, SEQ ID NO:12, SEQ ID NO:17 or SEQ ID NO:18.
 That is, variants can be obtained that contain one or more
 amino acid substitutions of SEQ ID NO:2, SEQ ID NO:3, SEQ
 ID NO: 4 SEQ ID NO:5 SEQ ID NO:9 or SEQ ID NO:12, in which
 an alkyl amino acid is substituted for an alkyl amino acid
 20 in a Ztgf β -9 amino acid sequence, an aromatic amino acid
 is substituted for an aromatic amino acid in an Ztgf β -9
 amino acid sequence, a sulfur-containing amino acid is
 substituted for a sulfur-containing amino acid in an
 Ztgf β -9 amino acid sequence, a hydroxy-containing amino
 25 acid is substituted for a hydroxy-containing amino acid in
 a Ztgf β -9 amino acid sequence, an acidic amino acid is
 substituted for an acidic amino acid in a Ztgf β -9 amino
 acid sequence, a basic amino acid is substituted for a
 basic amino acid in a Ztgf β -9 amino acid sequence, or a
 30 dibasic monocarboxylic amino acid is substituted for a
 dibasic monocarboxylic amino acid in an Ztgf β -9 amino acid
 sequence.

Among the common amino acids, for example, a
 35 "conservative amino acid substitution" is illustrated by a
 substitution among amino acids within each of the
 following groups: (1) glycine, alanine, valine, leucine,

and isoleucine, (2) phenylalanine, tyrosine, and tryptophan, (3) serine and threonine, (4) aspartate and glutamate, (5) glutamine and asparagine, and (6) lysine, arginine and histidine. For example, variant Ztgf β -9

5 polypeptides that have an amino acid sequence that differs from either SEQ ID NOs:2, 3, 4, 5 or 12 can be obtained by substituting a threonine residue for Ser, by substituting a valine residue for Ile, by substituting an aspartate residue for Glu, or by substituting a valine residue for

10 Ile. Additional variants can be obtained by producing polypeptides having two or more of these amino acid substitutions.

15 Variants of either the human or the murine Ztgf β -9 can be devised by aligning the amino acid sequences of SEQ ID NO:3 and SEQ ID NO:12, and by noting any differences in the corresponding amino acid residues.

The BLOSUM62 table is an amino acid substitution

20 matrix derived from about 2,000 local multiple alignments of protein sequence segments, representing highly conserved regions of more than 500 groups of related proteins (Henikoff and Henikoff, *Proc. Nat'l Acad. Sci. USA* 89:10915 (1992)). Accordingly, the BLOSUM62

25 substitution frequencies can be used to define conservative amino acid substitutions that may be introduced into the amino acid sequences of the present invention. Although it is possible to design amino acid substitutions based solely upon chemical properties (as

30 discussed above), the language "conservative amino acid substitution" preferably refers to a substitution represented by a BLOSUM62 value of greater than -1. For example, an amino acid substitution is conservative if the substitution is characterized by a BLOSUM62 value of 0,

35 1, 2, or 3. According to this system, preferred conservative amino acid substitutions are characterized by a BLOSUM62 value of at least 1 (e.g., 1, 2 or 3), while

more preferred conservative amino acid substitutions are characterized by a BLOSUM62 value of at least 2 (e.g., 2 or 3).

5 Particular variants of human or murine Ztgf β -9 are characterized by having at least 70%, at least 80%, at least 90%, at least 95% or 99% or greater sequence identity to the corresponding human (i.e., SEQ ID NOS: 2, 3, 4, 5 or 17) or murine (i.e., SEQ ID NOS:9 or 12) amino
10 acid sequences, wherein the variation in amino acid sequence is due to one or more conservative amino acid substitutions.

Conservative amino acid changes in a Ztgf β -9
15 gene can be introduced by substituting nucleotides for the nucleotides recited in any one of SEQ ID NOS:1 or 9. Such "conservative amino acid" variants can be obtained, for example, by oligonucleotide-directed mutagenesis, linker-scanning mutagenesis, mutagenesis using the polymerase
20 chain reaction, and the like (see Ausubel (1995) at pages 8-10 to 8-22; and McPherson (ed.), *Directed Mutagenesis: A Practical Approach* (IRL Press 1991)). The ability of such variants to promote anti-viral or anti-proliferative activity can be determined using a standard method, such
25 as the assay described herein. Alternatively, a variant Ztgf β -9 polypeptide can be identified by the ability to specifically bind anti- Ztgf β -9 antibodies.

The proteins of the present invention can also
30 comprise non-naturally occurring amino acid residues. Non-naturally occurring amino acids include, without limitation, *trans*-3-methylproline, 2,4-methanoproline, *cis*-4-hydroxyproline, *trans*-4-hydroxyproline, *N*-methylglycine, *allo*-threonine, methylthreonine,
35 hydroxyethylcysteine, hydroxyethylhomocysteine, nitroglutamine, homoglutamine, pipecolic acid, thiazolidine carboxylic acid, dehydroproline, 3- and 4-

methylethylproline, 3,3-dimethylethylproline, tert-leucine, norvaline, 2-azaphenylalanine, 3-azaphenylalanine, 4-azaphenylalanine, and 4-fluorophenylalanine. Several methods are known in the art for incorporating non-naturally occurring amino acid residues into proteins. For example, an *in vitro* system can be employed wherein nonsense mutations are suppressed using chemically aminoacylated suppressor tRNAs. Methods for synthesizing amino acids and aminoacylating tRNA are known in the art. Transcription and translation of plasmids containing nonsense mutations is typically carried out in a cell-free system comprising an *E. coli* S30 extract and commercially available enzymes and other reagents. Proteins are purified by chromatography. See, for example, Robertson et al., *J. Am. Chem. Soc.* 113:2722 (1991), Ellman et al., *Methods Enzymol.* 202:301 (1991), Chung et al., *Science* 259:806 (1993), and Chung et al., *Proc. Nat'l Acad. Sci. USA* 90:10145 (1993).

In a second method, translation is carried out in *Xenopus* oocytes by microinjection of mutated mRNA and chemically aminoacylated suppressor tRNAs (Turcatti et al., *J. Biol. Chem.* 271:19991 (1996)). Within a third method, *E. coli* cells are cultured in the absence of a natural amino acid that is to be replaced (e.g., phenylalanine) and in the presence of the desired non-naturally occurring amino acid(s) (e.g., 2-azaphenylalanine, 3-azaphenylalanine, 4-azaphenylalanine, or 4-fluorophenylalanine). The non-naturally occurring amino acid is incorporated into the protein in place of its natural counterpart. See, Koide et al., *Biochem.* 33:7470 (1994). Naturally occurring amino acid residues can be converted to non-naturally occurring species by *in vitro* chemical modification. Chemical modification can be combined with site-directed mutagenesis to further expand the range of substitutions (Wynn and Richards, *Protein Sci.* 2:395 (1993)).

A limited number of non-conservative amino acids, amino acids that are not encoded by the genetic code, non-naturally occurring amino acids, and unnatural amino acids may be substituted for Ztgf β -9 amino acid residues.

Essential amino acids in the polypeptides of the present invention can be identified according to procedures known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis (Cunningham and Wells, *Science* 244:1081 (1989), Bass et al., *Proc. Nat'l Acad. Sci. USA* 88:4498 (1991), Coombs and Corey, "Site-Directed Mutagenesis and Protein Engineering," in *Proteins: Analysis and Design*, Angeletti (ed.), pages 259-311 (Academic Press, Inc. 1998)). In the latter technique, single alanine mutations are introduced at every residue in the molecule, and the resultant mutant molecules are tested for biological activity as disclosed below to identify amino acid residues that are critical to the activity of the molecule. See also, Hilton et al., *J. Biol. Chem.* 271:4699 (1996).

Table 2

25	<u>Conservative amino acid substitutions</u>	
	Basic:	arginine
		lysine
		histidine
	Acidic:	glutamic acid
30		aspartic acid
	<u>Polar:</u>	glutamine
		asparagine
	Hydrophobic:	leucine
		isoleucine
35		valine
	Aromatic:	phenylalanine
		tryptophan

Small: tyrosine
glycine
alanine
serine
threonine
methionine

Essential amino acids in the polypeptides of the present invention can be identified according to procedures known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis [Cunningham and Wells, *Science* 244: 1081-1085 (1989); Bass et al., *Proc. Natl. Acad. Sci. USA* 88:4498-4502 (1991)]. In the latter technique, single alanine mutations are introduced at every residue in the molecule, and the resultant mutant molecules are tested for biological activity (e.g., ligand binding and signal transduction) to identify amino acid residues that are critical to the activity of the molecule. Sites of ligand-protein interaction can also be determined by analysis of crystal structure as determined by such techniques as nuclear magnetic resonance, crystallography or photoaffinity labeling. See, for example, de Vos et al., *Science* 255:306-312 (1992); Smith et al., *J. Mol. Biol.* 224:899-904, 1992; Wlodaver et al., *FEBS Lett.* 309:59-64 (1992). The identities of essential amino acids can also be inferred from analysis of homologies with related proteins.

Multiple amino acid substitutions can be made and tested using known methods of mutagenesis and screening, such as those disclosed by Reidhaar-Olson and Sauer, *Science* 241:53-57 (1988) or Bowie and Sauer, *Proc. Natl. Acad. Sci. USA* 86:2152-2156 (1989). Briefly, these authors disclose methods for simultaneously randomizing two or more positions in a polypeptide, selecting for functional polypeptide, and then sequencing the

mutagenized polypeptides to determine the spectrum of allowable substitutions at each position. Other methods that can be used include phage display, e.g., Lowman et al., *Biochem.* 30:10832-10837 (1991); Ladner et al., U.S. Patent No. 5,223,409; Huse, WIPO Publication WO 92/06204) and region-directed mutagenesis, Derbyshire et al., *Gene* 46:145 (1986); Ner et al., *DNA* 7:127 (1988).

Mutagenesis methods as disclosed above can be combined with high-throughput screening methods to detect activity of cloned, mutagenized proteins in host cells. Preferred assays in this regard include cell proliferation assays and biosensor-based ligand-binding assays, which are described below. Mutagenized DNA molecules that encode active proteins or portions thereof (e.g., ligand-binding fragments) can be recovered from the host cells and rapidly sequenced using modern equipment. These methods allow the rapid determination of the importance of individual amino acid residues in a polypeptide of interest, and can be applied to polypeptides of unknown structure.

Using the methods discussed above, one of ordinary skill in the art can prepare a variety of polypeptides that are substantially identical to SEQ ID NOs: 2, 3, 4, 5, 9, 12, 17 or 18 or allelic variants thereof and retain the properties of the wild-type protein. As expressed and claimed herein the language, "a polypeptide as defined by SEQ ID NOs: 2, 3, 4, 5, 9, 12, 17 or 18" includes all allelic variants and species orthologs of the polypeptide.

Another embodiment of the present invention provides for a peptide or polypeptide comprising an epitope-bearing portion of a polypeptide of the invention. The epitope of the this polypeptide portion is an immunogenic or antigenic epitope of a polypeptide of the

invention. A region of a protein to which an antibody can bind is defined as an "antigenic epitope". See for instance, Geysen, H.M. et al., *Proc. Natl. Acad. Sci. USA* 81:3998-4002 (1984).

5

As to the selection of peptides or polypeptides bearing an antigenic epitope (i.e., that contain a region of a protein molecule to which an antibody can bind), it is well known in the art that relatively short synthetic peptides that mimic part of a protein sequence are routinely capable of eliciting an antiserum that reacts with the partially mimicked protein. See Sutcliffe, J.G. et al. *Science* 219:660-666 (1983). Peptides capable of eliciting protein-reactive sera are frequently represented in the primary sequence of a protein, can be characterized by a set of simple chemical rules, and are confined neither to immunodominant regions of intact proteins (i.e., immunogenic epitopes) nor to the amino or carboxyl termini. Peptides that are extremely hydrophobic and those of six or fewer residues generally are ineffective at inducing antibodies that bind to the mimicked protein; longer soluble peptides, especially those containing proline residues, usually are effective.

25

Antigenic epitope-bearing peptides and polypeptides of the invention are therefore useful to raise antibodies, including monoclonal antibodies, that bind specifically to a polypeptide of the invention. Antigenic epitope-bearing peptides and polypeptides of the present invention contain a sequence of at least nine, preferably between 15 to about 30 amino acids contained within the amino acid sequence of a polypeptide of the invention. However, peptides or polypeptides comprising a larger portion of an amino acid sequence of the invention, containing from 30 to 50 amino acids, or any length up to and including the entire amino acid sequence of a polypeptide of the invention, also are useful for inducing

35

antibodies that react with the protein. Preferably, the amino acid sequence of the epitope-bearing peptide is selected to provide substantial solubility in aqueous solvents (*i.e.*, the sequence includes relatively

5 hydrophilic residues and hydrophobic residues are preferably avoided); and sequences containing proline residues are particularly preferred. All of the polypeptides shown in the sequence listing contain antigenic epitopes to be used according to the present

10 invention.

Polynucleotides, generally a cDNA sequence, of the present invention encode the above-described polypeptides. A cDNA sequence which encodes a polypeptide

15 of the present invention is comprised of a series of codons, each amino acid residue of the polypeptide being encoded by a codon and each codon being comprised of three nucleotides. The amino acid residues are encoded by their respective codons as follows.

20 Alanine (Ala) is encoded by GCA, GCC, GCG or GCT;

Cysteine (Cys) is encoded by TGC or TGT;

Aspartic acid (Asp) is encoded by GAC or GAT;

25 Glutamic acid (Glu) is encoded by GAA or GAG;

Phenylalanine (Phe) is encoded by TTC or TTT;

Glycine (Gly) is encoded by GGA, GGC, GGG or GGT;

Histidine (His) is encoded by CAC or CAT;

30 Isoleucine (Ile) is encoded by ATA, ATC or ATT;

Lysine (Lys) is encoded by AAA, or AAG;

Leucine (Leu) is encoded by TTA, TTG, CTA, CTC, CTG or CTT;

Methionine (Met) is encoded by ATG;

35 Asparagine (Asn) is encoded by AAC or AAT;

Proline (Pro) is encoded by CCA, CCC, CCG or CCT;

Glutamine (Gln) is encoded by CAA or CAG;
 Arginine (Arg) is encoded by AGA, AGG, CGA, CGC,
 CGG or CGT;

Serine (Ser) is encoded by AGC, AGT, TCA, TCC,
 5 TCG or TCT;

Threonine (Thr) is encoded by ACA, ACC, ACG or
 ACT;

Valine (Val) is encoded by GTA, GTC, GTG or GTT;

Tryptophan (Trp) is encoded by TGG; and

10 Tyrosine (Tyr) is encoded by TAC or TAT.

It is to be recognized that according to the
 present invention, when a cDNA is claimed as described
 above, it is understood that what is claimed are both the
 15 sense strand, the anti-sense strand, and the DNA as
 double-stranded having both the sense and anti-sense
 strand annealed together by their respective hydrogen
 bonds. Also claimed is the messenger RNA (mRNA) which
 encodes the polypeptides of the present invention, and
 20 which mRNA is encoded by the above-described cDNA. A
 messenger RNA (mRNA) will encode a polypeptide using the
 same codons as those defined above, with the exception
 that each thymine nucleotide (T) is replaced by a uracil
 nucleotide (U).

25

The protein polypeptides of the present
 invention, including full-length proteins, protein
 fragments (e.g. receptor-binding fragments), and fusion
 polypeptides can be produced in genetically engineered
 30 host cells according to conventional techniques. Suitable
 host cells are those cell types that can be transformed or
 transfected with exogenous DNA and grown in culture, and
 include bacteria, fungal cells, and cultured higher
 eukaryotic cells. Eukaryotic cells, particularly cultured
 35 cells of multicellular organisms, are preferred.
 Techniques for manipulating cloned DNA molecules and
 introducing exogenous DNA into a variety of host cells are

disclosed by Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, (2nd ed.) (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989).

5 In general, a DNA sequence encoding a Ztgf β -9 polypeptide is operably linked to other genetic elements required for its expression, generally including a transcription promoter and terminator, within an expression vector. The vector will also commonly contain
10 one or more selectable markers and one or more origins of replication, although those skilled in the art will recognize that within certain systems selectable markers may be provided on separate vectors, and replication of the exogenous DNA may be provided by integration into the
15 host cell genome. Selection of promoters, terminators, selectable markers, vectors and other elements is a matter of routine design within the level of ordinary skill in the art. Many such elements are described in the literature and are available through commercial suppliers.

20 To direct a Ztgf β -9 polypeptide into the secretory pathway of a host cell, a secretory signal sequence (also known as a leader sequence, prepro sequence or pre sequence) is provided in the expression vector.
25 The secretory signal sequence may be that of the protein, or may be derived from another secreted protein [e.g., the tissue plasminogen activator (t-PA)] leader sequence or synthesized *de novo*. The secretory signal sequence is joined to the Ztgf β -9 DNA sequence in the correct reading
30 frame. Secretory signal sequences are commonly positioned 5' to the DNA sequence encoding the polypeptide of interest, although certain signal sequences may be positioned elsewhere in the DNA sequence of interest (see, e.g., Welch *et al.*, U.S. Patent No. 5,037,743; Holland *et al.*, U.S. Patent No. 5,143,830).
35

Cultured mammalian cells are preferred hosts within the present invention. Methods for introducing exogenous DNA into mammalian host cells include calcium phosphate-mediated transfection, Wigler et al., *Cell* 5 14:725 (1978); Corsaro and Pearson, *Somatic Cell Genetics* 7:603 (1981); Graham and Van der Eb, *Virology* 52:456 (1973), electroporation, Neumann et al., *EMBO J.* 1:841-845 (1982), DEAE-dextran mediated transfection, Ausubel et al., eds., *Current Protocols in Molecular Biology* (John 10 Wiley and Sons, Inc., NY, 1987), and liposome-mediated transfection (Hawley-Nelson et al., *Focus* 15:73 (1993); Ciccarone et al., *Focus* 15:80 (1993)). The production of recombinant polypeptides in cultured mammalian cells is disclosed, for example, by Levinson et al., U.S. Patent 15 No. 4,713,339; Hagen et al., U.S. Patent No. 4,784,950; Palmiter et al., U.S. Patent No. 4,579,821; and Ringold, U.S. Patent No. 4,656,134. Suitable cultured mammalian cells include the COS-1 (ATCC No. CRL 1650), COS-7 (ATCC No. CRL 1651), BHK (ATCC No. CRL 1632), BHK 570 (ATCC No. 20 CRL 10314), 293 [ATCC No. CRL 1573; Graham et al., *J. Gen. Virol.* 36:59-72 (1977)] and Chinese hamster ovary (e.g. CHO-K1; ATCC No. CCL 61) cell lines. Additional suitable cell lines are known in the art and available from public depositories such as the American Type Culture Collection, 25 Rockville, Maryland. In general, strong transcription promoters are preferred, such as promoters from SV-40 or cytomegalovirus. See, e.g., U.S. Patent No. 4,956,288. Other suitable promoters include those from metallothionein genes (U.S. Patent Nos. 4,579,821 and 30 4,601,978) and the adenovirus major late promoter.

Other higher eukaryotic cells can also be used as hosts, including plant cells, insect cells and avian cells. The use of *Agrobacterium rhizogenes* as a vector 35 for expressing genes in plant cells has been reviewed by Sinkar et al., *J. Biosci. (Bangalore)* 11:47 (1987). Transformation of insect cells and production of foreign

polypeptides therein is disclosed by Guarino et al., U.S. Patent No. 5,162,222 and WIPO publication WO 94/06463. Insect cells can be infected with recombinant baculovirus, commonly derived from *Autographa californica* nuclear polyhedrosis virus (AcNPV). DNA encoding the Ztgf β -9

5 polypeptide is inserted into the baculoviral genome in place of the AcNPV polyhedrin gene coding sequence by one of two methods. The first is the traditional method of homologous DNA recombination between wild-type AcNPV and a

10 transfer vector containing the Ztgf β -9 cDNA flanked by AcNPV sequences. Suitable insect cells, e.g. SF9 cells, are infected with wild-type AcNPV and transfected with a transfer vector comprising a Ztgf β -9 polynucleotide operably linked to an AcNPV polyhedrin gene promoter,

15 terminator, and flanking sequences. See, King, L.A. and Possee, R.D., *The Baculovirus Expression System: A Laboratory Guide*, (Chapman & Hall, London); O'Reilly, D.R. et al., *Baculovirus Expression Vectors: A Laboratory Manual* (Oxford University Press, New York, New York,

20 1994); and, Richardson, C. D., Ed., *Baculovirus Expression Protocols. Methods in Molecular Biology*, (Humana Press, Totowa, NJ 1995). Natural recombination within an insect cell will result in a recombinant baculovirus which contains Ztgf β -9 driven by the polyhedrin promoter.

25 Recombinant viral stocks are made by methods commonly used in the art.

The second method of making recombinant baculovirus utilizes a transposon-based system described

30 by Luckow, V.A, et al., *J Virol* 67:4566 (1993). This system is sold in the Bac-to-Bac kit (Life Technologies, Rockville, MD). This system utilizes a transfer vector, pFastBac1TM (Life Technologies) containing a Tn7 transposon to move the DNA encoding the Ztgf β -9 polypeptide into a

35 baculovirus genome maintained in *E. coli* as a large plasmid called a "bacmid." The pFastBac1TM transfer vector

utilizes the AcNPV polyhedrin promoter to drive the expression of the gene of interest, in this case Ztgf β -9. However, pFastBac1™ can be modified to a considerable degree. The polyhedrin promoter can be removed and substituted with the baculovirus basic protein promoter (also known as Pcor, p6.9 or MP promoter) which is expressed earlier in the baculovirus infection, and has been shown to be advantageous for expressing secreted proteins. See, Hill-Perkins, M.S. and Possee, R.D., *J Gen Virol* 71:971 (1990); Bonning, B.C. et al., *J Gen Virol* 75:1551 (1994); and, Chazenbalk, G.D., and Rapoport, B., *J Biol Chem* 270:1543 (1995). In such transfer vector constructs, a short or long version of the basic protein promoter can be used. Moreover, transfer vectors can be constructed which replace the native Ztgf β -9 secretory signal sequences with secretory signal sequences derived from insect proteins. For example, a secretory signal sequence from Ecdysteroid Glucosyltransferase (EGT), honey bee Melittin (Invitrogen, Carlsbad, CA), or baculovirus gp67 (PharMingen, San Diego, CA) can be used in constructs to replace the native Ztgf β -9 secretory signal sequence. In addition, transfer vectors can include an in-frame fusion with DNA encoding an epitope tag at the C- or N-terminus of the expressed Ztgf β -9 polypeptide, for example, a Glu-Glu epitope tag, Grussenmeyer, T. et al., *Proc Natl Acad Sci.* 82:7952 (1985). Using a technique known in the art, a transfer vector containing Ztgf β -9 is transformed into *E. coli*, and screened for bacmids which contain an interrupted lacZ gene indicative of recombinant baculovirus. The bacmid DNA containing the recombinant baculovirus genome is isolated, using common techniques, and used to transfect *Spodoptera frugiperda* cells, e.g. Sf9 cells. Recombinant virus that expresses Ztgf β -9 is

subsequently produced. Recombinant viral stocks are made by methods commonly used the art.

The recombinant virus is used to infect host
5 cells, typically a cell line derived from the fall army
worm, *Spodoptera frugiperda*. See, in general, Glick and
Pasternak, *Molecular Biotechnology: Principles and
Applications of Recombinant DNA*, ASM Press, Washington,
D.C. (1994). Another suitable cell line is the High
10 FiveO™ cell line (Invitrogen) derived from *Trichoplusia ni*
(U.S. Patent #5,300,435). Commercially available serum-
free media are used to grow and maintain the cells.
Suitable media are Sf900 II™ (Life Technologies) or ESF
921™ (Expression Systems) for the Sf9 cells; and Ex-
15 cello405™ (JRH Biosciences, Lenexa, KS) or Express FiveO™
(Life Technologies) for the *T. ni* cells. The cells are
grown up from an inoculation density of approximately $2-5 \times 10^5$ cells to a density of $1-2 \times 10^6$ cells at which time a
recombinant viral stock is added at a multiplicity of
20 infection (MOI) of 0.1 to 10, more typically near 3. The
recombinant virus-infected cells typically produce the
recombinant Ztgfβ-9 polypeptide at 12-72 hours post-
infection and secrete it with varying efficiency into the
medium. The culture is usually harvested 48 hours post-
25 infection. Centrifugation is used to separate the cells
from the medium (supernatant). The supernatant containing
the z*** polypeptide is filtered through micropore
filters, usually 0.45 μm pore size. Procedures used are
generally described in available laboratory manuals (King,
30 L. A. and Pössee, R.D., *ibid.*; O'Reilly, D.R. et al.,
ibid.; Richardson, C. D., *ibid.*). Subsequent purification
of the Ztgfβ-9 polypeptide from the supernatant can be
achieved using methods described herein.

Drug selection is generally used to select for cultured mammalian cells into which foreign DNA has been inserted. Such cells are commonly referred to as "transfectants". Cells that have been cultured in the presence of the selective agent and are able to pass the gene of interest to their progeny are referred to as "stable transfectants." A preferred selectable marker is a gene encoding resistance to the antibiotic neomycin. Selection is carried out in the presence of a neomycin-type drug, such as G-418 or the like. Selection systems may also be used to increase the expression level of the gene of interest, a process referred to as "amplification." Amplification is carried out by culturing transfectants in the presence of a low level of the selective agent and then increasing the amount of selective agent to select for cells that produce high levels of the products of the introduced genes. A preferred amplifiable selectable marker is dihydrofolate reductase, which confers resistance to methotrexate. Other drug resistance genes (e.g. hygromycin resistance, multi-drug resistance, and puromycin acetyltransferase) can also be used.

Other higher eukaryotic cells can also be used as hosts, including insect cells, plant cells and avian cells. Transformation of insect cells and production of foreign polypeptides therein is disclosed by Guarino et al., U.S. Patent No. 5,162,222; Bang et al., U.S. Patent No. 4,775,624; and WIPO publication WO 94/06463. The use of *Agrobacterium rhizogenes* as a vector for expressing genes in plant cells has been reviewed by Sinkar et al., *J. Biosci. (Bangalore)* 11:47-58 (1987).

Fungal cells, including yeast cells, and particularly cells of the genus *Saccharomyces*, can also be used within the present invention, such as for producing protein fragments or polypeptide fusions. Methods for

transforming yeast cells with exogenous DNA and producing recombinant polypeptides therefrom are disclosed by, for example, Kawasaki, U.S. Patent No. 4,599,311; Kawasaki et al., U.S. Patent No. 4,931,373; Brake, U.S. Patent No. 4,870,008; Welch et al., U.S. Patent No. 5,037,743; and Murray et al., U.S. Patent No. 4,845,075. Transformed cells are selected by phenotype determined by the selectable marker, commonly drug resistance or the ability to grow in the absence of a particular nutrient (e.g., leucine). A preferred vector system for use in yeast is the *POT1* vector system disclosed by Kawasaki et al., U.S. Patent No. 4,931,373, which allows transformed cells to be selected by growth in glucose-containing media. Suitable promoters and terminators for use in yeast include those from glycolytic enzyme genes (see, e.g., Kawasaki, U.S. Patent No. 4,599,311; Kingsman et al., U.S. Patent No. 4,615,974; and Bitter, U.S. Patent No. 4,977,092) and alcohol dehydrogenase genes. See also U.S. Patents Nos. 4,990,446; 5,063,154; 5,139,936 and 4,661,454.

Transformation systems for other yeasts, including *Hansenula polymorpha*, *Schizosaccharomyces pombe*, *Kluyveromyces lactis*, *Kluyveromyces fragilis*, *Ustilago maydis*, *Pichia pastoris*, *Pichia methanolica*, *Pichia guilliermondii* and *Candida maltosa* are known in the art.

See, for example, Gleeson et al., *J. Gen. Microbiol.* 132:3459-3465 (1986) and Cregg, U.S. Patent No. 4,882,279. *Aspergillus* cells may be utilized according to the methods of McKnight et al., U.S. Patent No. 4,935,349. Methods for transforming *Acremonium chrysogenum* are disclosed by Sumino et al., U.S. Patent No. 5,162,228. Methods for transforming *Neurospora* are disclosed by Lambowitz, U.S. Patent No. 4,486,533.

Prokaryotic host cells, including strains of the bacteria *Escherichia coli*, *Bacillus* and other genera are also useful host cells within the present invention. Techniques for transforming these hosts and expressing

foreign DNA sequences cloned therein are well known in the art, see, e.g., Sambrook et al., *ibid.*). When expressing a Ztgf β -9 polypeptide in bacteria such as *E. coli*, the polypeptide may be retained in the cytoplasm, typically as insoluble granules, or may be directed to the periplasmic space by a bacterial secretion sequence. In the former case, the cells are lysed, and the granules are recovered and denatured using, for example, guanidine isothiocyanate or urea. The denatured polypeptide can then be refolded and dimerized by diluting the denaturant, such as by dialysis against a solution of urea and a combination of reduced and oxidized glutathione, followed by dialysis against a buffered saline solution. In the latter case, the polypeptide can be recovered from the periplasmic space in a soluble and functional form by disrupting the cells (by, for example, sonication or osmotic shock) to release the contents of the periplasmic space and recovering the protein, thereby obviating the need for denaturation and refolding.

Transformed or transfected host cells are cultured according to conventional procedures in a culture medium containing nutrients and other components required for the growth of the chosen host cells. A variety of suitable media, including defined media and complex media, are known in the art and generally include a carbon source, a nitrogen source, essential amino acids, vitamins and minerals. Media may also contain such components as growth factors or serum, as required. The growth medium will generally select for cells containing the exogenously added DNA by, for example, drug selection or deficiency in an essential nutrient which is complemented by the selectable marker carried on the expression vector or co-transfected into the host cell.

Within one aspect of the present invention, a novel protein is produced by a cultured cell, and the cell

or the protein is used to screen for a receptor or receptors for the protein, including the natural receptor, as well as interacting proteins such as dimerization partners, agonists and antagonists of the natural ligand.

5

PROTEIN ISOLATION:

Expressed recombinant polypeptides (or chimeric polypeptides) can be purified using fractionation and/or
10 conventional purification methods and media. Ammonium sulfate precipitation and acid or chaotrope extraction may be used for fractionation of samples. Exemplary purification steps may include hydroxyapatite, size exclusion, FPLC and reverse-phase high performance liquid
15 chromatography. Suitable anion exchange media include derivatized dextrans, agarose, cellulose, polyacrylamide, specialty silicas, and the like. PEI, DEAE, QAE and Q derivatives are preferred, with DEAE Fast-Flow Sepharose (Pharmacia, Piscataway, NJ) being particularly preferred.
20 Exemplary chromatographic media include those media derivatized with phenyl, butyl, or octyl groups, such as Phenyl-Sepharose FF (Pharmacia), Toyopearl butyl 650 (Toso Haas, Montgomeryville, PA), Octyl-Sepharose (Pharmacia) and the like; or polyacrylic resins, such as Amberchrom CG
25 71 (Toso Haas) and the like. Suitable solid supports include glass beads, silica-based resins, cellulosic resins, agarose beads, cross-linked agarose beads, polystyrene beads, cross-linked polyacrylamide resins and the like that are insoluble under the conditions in which
30 they are to be used. These supports may be modified with reactive groups that allow attachment of proteins by amino groups, carboxyl groups, sulfhydryl groups, hydroxyl groups and/or carbohydrate moieties. Examples of coupling chemistries include cyanogen bromide activation, N-
35 hydroxysuccinimide activation, epoxide activation, sulfhydryl activation, hydrazide activation, and carboxyl and amino derivatives for carbodiimide coupling

chemistries. These and other solid media are well known and widely used in the art, and are available from commercial suppliers. Methods for binding receptor polypeptides to support media are well known in the art.

- 5 Selection of a particular method is a matter of routine design and is determined in part by the properties of the chosen support. See, for example, *Affinity Chromatography: Principles & Methods* (Pharmacia LKB Biotechnology, Uppsala, Sweden, 1988).

10

- The polypeptides of the present invention can be isolated by exploitation of their *properties*. For example, immobilized metal ion adsorption (IMAC) chromatography can be used to purify histidine-rich
- 15 proteins. Briefly, a gel is first charged with divalent metal ions to form a chelate [E. Sulkowski, *Trends in Biochem.* 3:1-7 (1985)]. Histidine-rich proteins will be adsorbed to this matrix with differing affinities, depending upon the metal ion used, and will be eluted by
- 20 competitive elution, lowering the pH, or use of strong chelating agents. Other methods of purification include purification of glycosylated proteins by lectin affinity chromatography and ion exchange chromatography [*Methods in Enzymol.*, Vol. 182:529-39, "Guide to Protein
- 25 Purification", M. Deutscher, (ed.), (Acad. Press, San Diego, 1990). Alternatively, a fusion of the polypeptide of interest and an affinity tag (e.g., polyhistidine, maltose-binding protein, an immunoglobulin domain) may be constructed to facilitate purification. Furthermore, to
- 30 facilitate purification of the secreted polypeptide, an amino or carboxyl-terminal extension, such as a polyhistidine tag, substance P, FLAG[®] peptide [Hopp et al., *Bio/Technology* 6:1204-1210 (1988); available from Eastman Kodak Co., New Haven, CT), a Glu-Glu affinity tag
- 35 [Grussenmeyer et al., *Proc. Natl. Acad. Sci. USA* 82:7952-4 (1985)], or another polypeptide or protein for which an

antibody or other specific binding agent is available, can be fused to Ztgfb β to aid in purification.

Mice engineered to express the Ztgfb β -9 gene, referred to as "transgenic mice" and mice that exhibit a complete absence of Ztgfb β -9 gene function, referred to as "knockout mice", may also be generated (Snouwaert *et al.*, *Science*, 257:1083 (1992); Lowell, *et al.*, *Nature*, 366:740-742 (1993); Capecchi, M.R., *Science*, 244:1288-1292, (1989); Palmiter, R.D. *et al.*, *Annu. Rev. Genet.*, 20:465-499, (1986). For example, transgenic mice that over-express Ztgfb β -9 either ubiquitously or under a tissue-specific or tissue-restricted promoter can be used to ask whether overexpression causes a phenotype. For example, overexpression of a wild-type Ztgfb β -9 polypeptide, polypeptide fragment or a mutant thereof may alter normal cellular processes resulting in a phenotype that identifies a tissue in which Ztgfb β -9 expression is functionally relevant and may indicate a therapeutic target for the Ztgfb β -9 protein, gene, its agonists or antagonists. Moreover, such over-expression may result in a phenotype that shows similarity with human diseases. Similarly, knockout Ztgfb β -9 mice can be used to determine where Ztgfb β -9 is absolutely required *in vivo*. The phenotype of knockout mice is predictive of the *in vivo* effects of that of a Ztgfb β -9 antagonist. The human Ztgfb β -9 cDNA can be used to isolate murine Ztgfb β -9 mRNA, cDNA and genomic DNA, which are subsequently used to generate knockout or transgenic mice. These mice may be employed to study the Ztgfb β -9 gene and the protein encoded thereby in an *in vivo* system, and can be used as *in vivo* models for corresponding human diseases. Moreover, transgenic mouse expression of Ztgfb β -9 antisense polynucleotides or ribozymes directed against Ztgfb β -9 or single chain antibodies to Ztgfb β -9 can be used to further elucidate the biology of Ztgfb β -9.

Uses

Northern blot analysis of the expression of Ztgfb β -9 reveals that Ztgfb β -9 is highly expressed in the brain and spinal cord. Therefore, Ztgfb β -9 may play a role in the maintenance of spinal cord involving either glial cells or neurons. This indicates that Ztgfb β -9 can be used to treat a variety of neurodegenerative diseases such as amyotrophic lateral sclerosis (ALS), Alzheimer's disease, Huntington's disease, Parkinson's disease and peripheral neuropathies, or demyelinating diseases including multiple sclerosis. The tissue specificity of Ztgfb β -9 expression suggests that Ztgfb β -9 may be a growth and/or maintenance factor in the spinal cord and brain which can be used to treat spinal cord, brain or peripheral nervous system injuries. Ztgfb β -9 can also be administered to someone to treat a viral infection.

The present invention also provides reagents with significant therapeutic value. The Ztgfb β -9 polypeptide (naturally occurring or recombinant), fragments thereof, antibodies and anti-idiotypic antibodies thereto, along with compounds identified as having binding affinity to the Ztgfb β -9 polypeptide, should be useful in the treatment of conditions associated with abnormal physiology or development, including abnormal proliferation, e.g., cancerous conditions, or degenerative conditions. For example, a disease or disorder associated with abnormal expression or abnormal signaling by a Ztgfb β -9 polypeptide should be a likely target for an agonist or antagonist of the Ztgfb β -9 polypeptide. In particular, Ztgfb β -9 can be used to treat inflammation. Inflammation is a result of an immune response to an infection or as an autoimmune response to a self-antigen.

35

Antibodies to the Ztgfb β -9 polypeptide can be purified and then administered to a patient. These

reagents can be combined for therapeutic use with additional active or inert ingredients, e.g., in pharmaceutically acceptable carriers or diluents along with physiologically innocuous stabilizers and excipients.

5 These combinations can be sterile filtered and placed into dosage forms as by lyophilization in dosage vials or storage in stabilized aqueous preparations. This invention also contemplates use of antibodies, binding fragments thereof or single-chain antibodies of the antibodies

10 including forms which are not complement binding.

The quantities of reagents necessary for effective therapy will depend upon many different factors, including means of administration, target site,

15 physiological state of the patient, and other medications administered. Thus, treatment dosages should be titrated to optimize safety and efficacy. Typically, dosages used *in vitro* may provide useful guidance in the amounts useful for *in vivo* administration of these reagents. Animal

20 testing of effective doses for treatment of particular disorders will provide further predictive indication of human dosage. Methods for administration include oral, intravenous, peritoneal, intramuscular, or transdermal administration. Pharmaceutically acceptable carriers will

25 include water, saline or buffers to name just a few. Dosage ranges would ordinarily be expected from 1 μ g to 1000 μ g per kilogram of body weight per day. However, the doses may be higher or lower as can be determined by a medical doctor with ordinary skill in the art. For a

30 complete discussion of drug formulations and dosage ranges see *Remington's Pharmaceutical Sciences*, 17th Ed., (Mack Publishing Co., Easton, Penn., 1990), and *Goodman and Gilman's: The Pharmacological Bases of Therapeutics*, 9th Ed. (Pergamon Press 1996).

Nucleic Acid-based Therapeutic Treatment

If a mammal has a mutated or lacks a *Ztgfβ-9* gene, the *Ztgfβ-9* gene can be introduced into the cells of the mammal. In one embodiment, a gene encoding a *Ztgfβ-9* polypeptide is introduced *in vivo* in a viral vector. Such vectors include an attenuated or defective DNA virus, such as but not limited to herpes simplex virus (HSV), papillomavirus, Epstein Barr virus (EBV), adenovirus, adeno-associated virus (AAV), SV40 and the like. Defective viruses, which entirely or almost entirely lack viral genes, are preferred. A defective virus is not infective after introduction into a cell. Use of defective viral vectors allows for administration to cells in a specific, localized area, without concern that the vector can infect other cells. Examples of particular vectors include, but are not limited to, a defective herpes virus 1 (HSV1) vector [Kaplitt et al., *Molec. Cell. Neurosci.*, 2 :320-330 (1991)], an attenuated adenovirus vector, such as the vector described by Stratford-Perricaudet et al., *J. Clin. Invest.*, 90 :626-630 (1992), and a defective adeno-associated virus vector [Samulski et al., *J. Virol.*, 61:3096-3101 (1987); Samulski et al. *J. Virol.*, 63:3822-3828 (1989)].

25

In another embodiment, the gene can be introduced in a retroviral vector, e.g., as described in Anderson et al., U.S. Patent No. 5,399,346; Mann et al., *Cell*, 33:153 (1983); Temin et al., U.S. Patent No. 4,650,764; Temin et al., U.S. Patent No. 4,980,289; Markowitz et al., *J. Virol.*, 62:1120 (1988); Temin et al., U.S. Patent No. 5,124,263; International Patent Publication No. WO 95/07358, published March 16, 1995 by Dougherty et al.; and *Blood*, 82:845 (1993).

35

Alternatively, the vector can be introduced by lipofection *in vivo* using liposomes. Synthetic cationic

lipids can be used to prepare liposomes for *in vivo* transfection of a gene encoding a marker [Felgner et al., *Proc. Natl. Acad. Sci. USA*, 84:7413-7417 (1987); see Mackey et al., *Proc. Natl. Acad. Sci. USA*, 85:8027-8031 (1988)]. The use of lipofection to introduce exogenous genes into specific organs *in vivo* has certain practical advantages. Molecular targeting of liposomes to specific cells represents one area of benefit. It is clear that directing transfection to particular cell types would be particularly advantageous in a tissue with cellular heterogeneity, such as the pancreas, liver, kidney, and brain. Lipids may be chemically coupled to other molecules for the purpose of targeting. Targeted peptides, e.g., hormones or neurotransmitters, and proteins such as antibodies, or non-peptide molecules could be coupled to liposomes chemically.

It is possible to remove the cells from the body and introduce the vector as a naked DNA plasmid or by means of a viral vector and then re-implant the transformed cells into the body. Naked DNA vector for gene therapy can be introduced into the desired host cells by methods known in the art, e.g., transfection, electroporation, microinjection, transduction, cell fusion, DEAE dextran, calcium phosphate precipitation, use of a gene gun or use of a DNA vector transporter [see, e.g., Wu et al., *J. Biol. Chem.*, 267:963-967 (1992); Wu et al., *J. Biol. Chem.*, 263:14621-14624 (1988)]. Techniques such as viral vector-mediated gene delivery of Ztgf β -9 can be used to treat human diseases such as cancer, immune & autoimmune diseases, and diseases of the central and peripheral nervous system.

Ztgf β -9 polypeptides can also be used to prepare antibodies that specifically bind to Ztgf β -9 polypeptides. These antibodies can then be used to manufacture anti-idiotypic antibodies. As used herein, the term

"antibodies" includes polyclonal antibodies, monoclonal antibodies, antigen-binding fragments thereof such as $F(ab')_2$ and Fab fragments, and the like, including genetically engineered antibodies. Antibodies are defined to be specifically binding if they bind to a Ztgf β -9 polypeptide with a K_a of greater than or equal to $10^7/M$ and they do not substantially bind to a polypeptide of the prior art. The affinity of a monoclonal antibody can be readily determined by one of ordinary skill in the art, for example, by using Scatchard analysis.

Methods for preparing polyclonal and monoclonal antibodies are well known in the art (see for example, Sambrook et al., *Molecular Cloning: A Laboratory Manual*, (Second Edition) (Cold Spring Harbor, NY, 1989); and Hurrell, J. G. R., Ed., *Monoclonal Hybridoma Antibodies: Techniques and Applications* (CRC Press, Inc., Boca Raton, FL, 1982). Polyclonal antibodies can be generated by inoculating a variety of warm-blooded animals such as horses, cows, goats, sheep, dogs, chickens, rabbits, mice, hamsters, guinea pigs and rats with a Ztgf β -9 polypeptide or a fragment thereof. The immunogenicity of a Ztgf β -9 polypeptide may be increased through the use of an adjuvant, such as alum (aluminum hydroxide) or Freund's complete or incomplete adjuvant. Polypeptides useful for immunization also include fusion polypeptides, such as fusions of Ztgf β -9 or a portion thereof with an immunoglobulin polypeptide or with maltose binding protein. The polypeptide immunogen may be a full-length molecule or a portion thereof. If the polypeptide portion is "hapten-like", such portion may be advantageously joined or linked to a macromolecular carrier (such as keyhole limpet hemocyanin (KLH), bovine serum albumin (BSA) or tetanus toxoid) for immunization. A variety of assays known to those skilled in the art can be utilized to detect antibodies which specifically bind to Ztgf β -9 polypeptides. Exemplary assays are described in detail in

Antibodies: A Laboratory Manual, Harlow and Lane (Eds.), (Cold Spring Harbor Laboratory Press, 1988).

Representative examples of such assays include: concurrent immunoelectrophoresis, radio-immunoassays, radio-
5 immunoprecipitations, enzyme-linked immunosorbent assays (ELISA), dot blot assays, inhibition or competition assays, and sandwich assays.

As used herein, the term "antibodies" includes
10 polyclonal antibodies, affinity-purified polyclonal antibodies, monoclonal antibodies, and antigen-binding fragments, such as F(ab')₂ and Fab proteolytic fragments. Genetically engineered intact antibodies or fragments, such as chimeric antibodies, Fv fragments, single chain
15 antibodies and the like, as well as synthetic antigen-binding peptides and polypeptides, are also included. Non-human antibodies may be humanized by grafting non-human CDRs onto human framework and constant regions, or by incorporating the entire non-human variable domains
20 (optionally "cloaking" them with a human-like surface by replacement of exposed residues, wherein the result is a "veneered" antibody). In some instances, humanized antibodies may retain non-human residues within the human variable region framework domains to enhance proper
25 binding characteristics. Through humanizing antibodies, biological half-life may be increased, and the potential for adverse immune reactions upon administration to humans is reduced. Human antibodies can be generated in mice engineered to contain the human immunoglobulin loci,
30 Vaughan, et al. *Nat. Biotech.*, 16:535-539 (1998).

Alternative techniques for generating or selecting antibodies useful herein include *in vitro* exposure of lymphocytes to Ztgfβ-9 protein or peptide, and
35 selection of antibody display libraries in phage or similar vectors (for instance, through use of immobilized

or labeled Ztgfb-9 protein or peptide). Genes encoding polypeptides having potential Ztgfb-9 polypeptide binding domains can be obtained by screening random peptide libraries displayed on phage (phage display) or on
5 bacteria, such as *E. coli*. Nucleotide sequences encoding the polypeptides can be obtained in a number of ways, such as through random mutagenesis and random polynucleotide synthesis. These random peptide display libraries can be used to screen for peptides which interact with a known
10 target which can be a protein or polypeptide, such as a ligand or receptor, a biological or synthetic macromolecule, or organic or inorganic substances. Techniques for creating and screening such random peptide display libraries are known in the art (Ladner et al., US
15 Patent NO. 5,223,409; Ladner et al., US Patent NO. 4,946,778; Ladner et al., US Patent NO. 5,403,484 and Ladner et al., US Patent NO. 5,571,698) and random peptide display libraries and kits for screening such libraries are available commercially, for instance from Clontech
20 (Palo Alto, CA), Invitrogen Inc. (San Diego, CA), New England Biolabs, Inc. (Beverly, MA) and Pharmacia LKB Biotechnology Inc. (Piscataway, NJ). Random peptide display libraries can be screened using the Ztgfb-9 sequences disclosed herein to identify proteins which bind
25 to Ztgfb-9. These "binding proteins" which interact with Ztgfb-9 polypeptides can be used for tagging cells; for isolating homolog polypeptides by affinity purification; they can be directly or indirectly conjugated to drugs, toxins, radionuclides and the like. These binding
30 proteins can also be used in analytical methods such as for screening expression libraries and neutralizing activity. The binding proteins can also be used for diagnostic assays for determining circulating levels of polypeptides; for detecting or quantitating soluble
35 polypeptides as marker of underlying pathology or disease. These binding proteins can also act as Ztgfb-9

"antagonists" to block Ztgf β -9 binding and signal transduction *in vitro* and *in vivo*.

Antibodies can also be generated by gene therapy. The animal is administered the DNA or RNA which encodes Ztgf β -9 or an immunogenic fragment thereof so that cells of the animals are transfected with the nucleic acid and express the protein which in turn elicits an immunogenic response. Antibodies which then are produced by the animal are isolated in the form of polyclonal or monoclonal antibodies. Antibodies to Ztgf β -9 may be used for tagging cells that express the protein, for affinity purification, within diagnostic assays for determining circulating levels of soluble protein polypeptides, and as antagonists to block ligand binding and signal transduction *in vitro* and *in vivo*.

Radiation hybrid mapping is a somatic cell genetic technique developed for constructing high-resolution, contiguous maps of mammalian chromosomes [Cox et al., *Science* 250:245-250 (1990)]. Partial or full knowledge of a gene's sequence allows the designing of PCR primers suitable for use with chromosomal radiation hybrid mapping panels. Commercially available radiation hybrid mapping panels which cover the entire human genome, such as the Stanford G3 RH Panel and the GeneBridge 4 RH Panel (Research Genetics, Inc., Huntsville, AL), are available. These panels enable rapid, PCR based, chromosomal localizations and ordering of genes, sequence-tagged sites (STSs), and other nonpolymorphic- and polymorphic markers within a region of interest. This includes establishing directly proportional physical distances between newly discovered genes of interest and previously mapped markers. The precise knowledge of a gene's position can be useful in a number of ways including: 1) determining if a sequence is part of an existing contig and obtaining

additional surrounding genetic sequences in various forms such as genomic YAC-, BAC- or phage genomic clones or cDNA clones, 2) providing a possible candidate gene for an inheritable disease which shows linkage to the same
5 chromosomal region, and 3) for cross-referencing model organisms such as mouse which may be beneficial in helping to determine what function a particular gene might have.

The present invention also provides reagents
10 which will find use in diagnostic applications. For example, the *Ztgfβ-9* gene has been mapped on chromosome 13q11.2-q11. A *Ztgfβ-9* nucleic acid probe could be used to check for abnormalities on chromosome 13. For example, a probe comprising *Ztgfβ-9* DNA or RNA or a subsequence
15 thereof can be used to determine if the *Ztgfβ-9* gene is present on human chromosome 13q11.2-q11 or if a mutation has occurred. Detectable chromosomal aberrations at the *Ztgfβ-9* gene locus include but are not limited to aneuploidy, gene copy number changes, insertions,
20 deletions, restriction site changes and rearrangements. Such aberrations can be detected using polynucleotides of the present invention by employing molecular genetic techniques, such as restriction fragment length polymorphism (RFLP) analysis, short tandem repeat (STR)
25 analysis employing PCR techniques, and other genetic linkage analysis techniques known in the art. Human *Ztgfβ-9* maps at the 13q11.2-q11 region. Mouse *Ztgfβ-9* maps to mouse chromosome 14 framework markers d14mit64 and dmit82 located at 22.0 and 19.5 centimorgans, respectively. The
30 19.5 cm region appears to be syntenic with the human locus containing the gap junction genes *gja3* and *gjb2*. See Mignon, C. et al., *Cytogenet. Cell Genet.* 72: 185-186 (1996).

35 The invention is further illustrated by the following non-limiting examples.

Example 1Cloning of *Ztgfβ-9*

5 Human *Ztgfβ-9* was isolated from an arrayed
pituitary gland cDNA plasmid library by PCR screening
using SEQ ID NOs: 6 and 7. Thermocycler conditions were as
follows: one cycle at 94°C for 3 minutes, 35 cycles at 94°C
for 30 seconds, 62°C for 20 seconds, 72°C for 30 seconds,
10 one cycle at 72°C for 5 minutes, followed by 4° C hold. The
reactions were gel electrophoresed to identify positive
pools and, in this way the library was deconvoluted to a
pool of positive clones. These were electroporated into *E.*
coli DH10B cells and plated for colony hybridization. The
15 colonies were transferred to Hybond N filters (Amersham)
and probed for positive colonies. Positive clones were
sequenced for full length *Ztgfβ-9*.

Sequence analysis and conceptual translation of
20 the human *Ztgfβ-9* cDNA (SEQ ID NOs: 1 and 16) predicts a
protein product that is 202 amino acid residues in length.
This protein is homologous to two members of the IL-17
family, Zcyto7, International Application No.
PCT/US98/08212, and IL-17. *Ztgfβ-9* shares 27.8% amino acid
25 identity with Zcyto7 and 20.6% identity with IL-17 as
determined by the Clustal Method using Lasergene MegAlign
software. See Higgins and Sharp, *CABIOS* 5:151 (1989). In
particular, *Ztgfβ-9* shares four conserved cysteines (amino
acid residues 114, 119, 167, 169 in SEQ ID NO:2) with
30 Zcyto7 and IL-17. These cysteines are predicted to be
involved in forming a cysteine-knot-like protein fold that
is related to the found in TGF-β proteins.

Example 2Northern Analysis *Ztgfβ-9*

5 Analysis of tissue distribution was performed by
the northern blotting technique using 2 adult and 1 fetal
human brain blots, Human Multiple Tissue and Master Dot
Blots from Clontech (Palo Alto, Ca). A probe was
obtained by PCR using SEQ ID NOs:6 and 7. in a cDNA pool.
10 Thermocycler conditions were as follows: one cycle at 94°C
for 3 minutes, 35 cycles at 94°C for 10 seconds, 66°C for
20 seconds, 72°C for 30 seconds, one cycle at 72°C for 5
minutes, followed by 4°C hold. The reaction mixture was
electrophoresed on a preparative agarose gel and a 162 bp
15 fragment was gel purified using commercially available gel
purification reagents and protocol (QIAEX II Gel
Extraction Kit; Qiagen, Inc., Santa Clarita, Ca). The
purified DNA was radioactively labeled with ³²P using a
commercially available kit (Rediprime DNA labeling
20 system; Amersham Corp., Arlington Heights, IL). The probe
was purified using a NUCTRAP push column (Stratagene
Cloning Systems, La Jolla, CA) . EXPRESSHYB (Clontech,
Palo Alto, CA) solution was used for prehybridization and
hybridization. The hybridization solution consisted of 8
25 mls EXPRESSHYB, 80μl Sheared Salmon Sperm DNA (10mg/ml, 5
Prime-3 Prime, Boulder, CO), 48 μl Human Cot-1 DNA
(1mg/ml, GibcoBRL) and 18μl of radiolabeled probe.
Hybridization took place overnight at 50°C And the blots
were then washed in 2X SSC, 0.1% SDS at RT, then 2X SSC, 0.1%
30 SDS at 60°C, followed by 0.1X SSC, 0.1% SDS wash at 60°C.
The blots were exposed overnight and developed. A major
transcript signal of was observed on MTN blots in brain
and spinal cord.

35 Master Dot blot signals were strong in all brain
tissues (adult and fetal), spinal cord, heart, skeletal
muscle, stomach, pancreas, adrenal gland, salivary gland,

liver, small intestine, bone marrow, thymus, spleen, lymph node, heart, thyroid, trachea, testis, ovary and placenta.

Example 3

5

Cloning of Murine *Ztgfb-9*

Full length sequence was obtained from a clone isolated from an arrayed mouse testis cDNA/plasmid library. The library was screened by PCR using oligonucleotides SEQ NO:10 and SEQ ID NO:11. The library was deconvoluted down to a positive pool of 250 clones. *E.coli* DH10B cells (Gibco BRL) were transformed with this pool by electroporation. The transformed culture was
10 titered and arrayed out to 96 wells at ~20 cells/well. The cells were grown up in LBamp overnight at 37°C. An aliquot of the cells were pelleted and PCR was used to identify a positive pool. The remaining cells from a positive pool were plated and colonies screened by PCR to identify a
15 positive clone. The clone was sequenced and contained the putative full length sequence of murine *Ztgfb-9*. The sequence of murine *Ztgfbeta-9* is defined by SEQ ID NOs: 8 and 9.

25

Example 4

Northern Analysis of Mouse *Ztgfb-9*

Northern analysis was also carried out on Mouse
30 MTN and Master Dot blots (Clontech) and a Mouse Embryo blot. A full-length murine *Ztgfb-9* cDNA clone (see cloning section) was restriction digested with *ApaI* and *EcoRI* following standard protocols. The reaction was gel electrophoresed and the ~686bp fragment was gel purified
35 using the Qiaex II Gel Purification Kit (Qiagen, Valencia, CA). The cDNA was P32-labeled using the Rediprime II Labeling Kit (Amersham) and column purified using reagents

and protocols described earlier. Hybridization, washing, and detection were carried out under conditions as described in Example 2. A band was observed in heart, brain, lung, liver, skeletal muscle, kidney and testis.

- 5 The Master Dot blot had strong signals in thyroid, with fainter signals in most other tissues. Hybridization to the Mouse Embryo blot indicated that *Ztgf β -9* was expressed at all stages examined (embryonic days 7, 11, 15, and 17).

- 10 By quantitative RT-PCR, murine *Ztgf β -9* was found to be highly expressed in the HCL hypothalamic cell line, and at lower levels in the GT1-1 and GT1-7 hypothalamic cell lines and the undifferentiated P19 teratocarcinoma cell line. Using quantitative RT-PCR, murine *Ztgf β -9* was
15 detected in neurons of the hippocampal cerebellar and olfactory cortex, Purkinje cells and other neuronal populations were heavily labeled in brain sections. The endothelium of the choroid plexus was also heavily
20 positive. In the spinal cord, labeling was confined to the gray matter and appeared to be uniformly found in dorsal and ventral horn neurons representing sensory and motor neurons. Strong expression was also observed in the dorsal root ganglia.

Example 5

Antibody Production

5 Polypeptides SEQ ID NOs: 13, 14 and 15 were
synthesized and were injected into rabbits and polyclonal
anti-sera was subsequently affinity purified by column
chromatography using the cognate immunogen. Also fusion
proteins between full-length human and mouse Ztgf- β fused
10 to the C-terminus of the maltose-binding protein were
expressed in *E. coli* and purified by affinity
chromatography over an amylose resin. Purified proteins
were injected into rabbits and polyclonal anti-sera was
subsequently affinity purified by column chromatography
15 using the cognate immunogen.

Example 6

Immunocytochemistry

20 Affinity purified polyclonal antibodies to human
Ztgf β -9 produced according to the procedure of Example 5
were validated on normal COS cells and COS cells
transfected with a Ztgf β -9 mammalian cell expression
25 construct. Immunocytochemistry performed with anti-Ztgf β -9
antibodies demonstrated Ztgf β -9 expression in monkey brain
and spinal cord. The immunocytochemistry staining was
intracytoplasmic and observed in many large neurons and
Purkinje cells. Scattered epithelial cells in the human
30 duodenum also showed positive staining.

Example 7

Mammalian Cell Protein Production

35 Human Ztgf β -9 protein, both with and without a
C-terminal a Glu-Glu affinity tag [Grussenmeyer et al.,

Proc. Natl. Acad. Sci. USA 82:7952-4 (1985)], was expressed in BHK cells using an expression vector in which Ztgf β -9 expression is driven by the CMV immediate early promoter, a consensus intron from the variable region of mouse immunoglobulin heavy chain locus, multiple restriction sites for insertion of coding sequences, a stop codon and a human growth hormone terminator. The plasmid also has an *E. coli* origin of replication, a mammalian selectable marker expression unit having an SV40 promoter, enhancer and origin of replication, a DHFR gene and the SV40 terminator and Kozac sequences at the 5' end of the open reading frame.

Following selection for stable cell transfectants, media isolated from pools was analyzed by western blot under reducing and non-reducing conditions using a Ztgf β -9 antibody or an anti-EE epitope tag antibody. Human Ztgf β -9 was found to migrate under reducing conditions at 29 kDa. Since the predicted molecular weight of the fully processed form of the protein is 20.31 kDa, this suggests that the protein is glycosylated at one or both of two potential glycosylation sites. Under non-reducing conditions, Ztgf β -9 protein migrated as at 49 kDa species. These results indicate that human Ztgf β -9 is capable of forming a disulfide cross-linked homodimer. However, co-expression of C-terminally EE tagged human Ztgf β -9 and untagged human Zcyto7, followed by affinity purification using an anti-EE tag antibody resulted in co-purification of both proteins, suggesting that in addition to Ztgf β -9 homodimers, Ztgf β -9 and Zcyto7 can also dimerize. Interaction of Ztgf β -9 and Zcyto7 did not appear to be due to interchain disulfide-bonding between the proteins. C-terminal EE tagged human Ztgf β -9 expressed in BHK cells was also anti-EE affinity purified and its N-terminal sequence determined. The signal cleavage site was found to occur proceeding amino acid A23.

Example 8

Transgenic Mice

5

The open reading frame encoding full-length murine Ztgfb-9 was amplified by PCR so as to introduce an optimized initiation codon and introduced into a transgenic vector in which expression of Ztgfb-9 was regulated by the metallothionein I promoter. The transgene insert was separated from plasmid backbone by NotI digestion and agarose gel purification, and fertilized ova from matings of B6C3F1Tac mice were microinjected and implanted into pseudopregnant females. Founders were identified by PCR on genomic tail DNA (DNAeasy 96 kit; Qiagen). Transgenic lines were initiated by breeding founders with C57BL/6Tac mice. Animal protocols used in this study were approved by the ZymoGenetics Institutional Animal Care and Use Committee. From 49 progeny born only 8% were found to be transgenic (compared to an average of 20% observed for a variety of other cDNAs driven by the same promoter), suggesting that high expression of murine Ztgfb-9 may be embryonic lethal. Consistent with this, of the four founders identified, all expressed only low levels of Ztgfb-9 mRNA in the liver. A fifth founder died at birth and interestingly, this animal expressed very high levels of Ztgfb-9 mRNA in the liver (8500 copies/cell). Histopathological analysis of this animal identified severe apoptosis of the thymus and complete devaculization of brown fat. The expressing males were bred with wild-type females. One founder was capable of germline transmission, however all transgenic progeny of this founder either died at birth or were runted and died soon after weaning. Analysis of these animals identified a variety of phenotypes, including severe thymic apoptosis, devaculization of brown fat, liver hepatitis, and low

lymphocyte peripheral blood cell numbers. These results indicate that Ztgfb-9, its agonists and antagonists, and antibodies to Ztgfb-9 will be useful in regulating immune cells, adipogenesis, and liver cells.

5

Example 9

Chromosomal Position

10 Human Ztgfb-9 was mapped to chromosome 13q11.2 on two radiation hybrid panels. The mouse Ztgfb9 gene links to murine chromosome 14 framework markers d14mit64 and dmit82 located at 22.0 and 19.5 centimorgans, respectively.

15

Example 10

Inhibition of Adenovirus Growth by Ztgfb-9

20 The human and murine Ztgfb-9 coding region were cloned into the adenovirus shuttle vector and recombined to generated the recombinant adenoviral genome. Transfection of Ztgfb-9 adenoviral genomes into 293A cells resulted in very small viral plaques (1 or 2 plaques per
25 transfection which is low). These plaques did not expand in size. Normally plaques expand greatly in size over a period of 1-2 days as the virus replicates. The small plaques were harvested and we tried to expand the virus by infecting 293A cells. Infected monolayers again exhibited
30 very low numbers of plaques and the plaques were small in size. These plaques did not expand in size over time. After 2-3 rounds of attempting to amplify the virus, eventually we did obtain a rapidly growing virus population. The virus that results from this
35 amplification still contains the Ztgfb-9 sequences. However, infection of cell by these viruses did not result in protein production. The initial behavior of both the

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